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FATTY ACIDS

(57) Abstract

By this invention, compositions and methods of use related to β -ketoacyl-ACP synthase of special interest are synthases obtainable from Cuphea species. Amino acid and nucleic acid for synthase protein factors are provided, as well as methods to utilize such sequences in constructs for production of genetically engineered plants having altered fatty acid compositions. Of particular interest is the expression of synthase protein factors in conjunction with expression of plant medium-chain acyl-ACP thioesterases for production of increased levels and/or modified ratios of medium-chain fatty acids in oils of transgenic plant seeds.

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PLANT FATTY ACID SYNTHASES AND USE IN IMPROVED METHODS FOR PRODUCTION OF MEDIUM-CHAIN FATTY ACIDS

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INTRODUCTION

Field of Invention

The present invention is directed to genes encoding

plant fatty acid synthase enzymes relevant to fatty acid

synthesis in plants, and to methods of using such genes in

combination with genes encoding plant medium-chain

preferring thioesterase proteins. Such uses provide a

method to increase the levels of medium-chain fatty acids

that may be produced in seed oils of transgenic plants.

Background

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Higher plants synthesize fatty acids via a common metabolic pathway. In developing seeds, where fatty acids attached to triglycerides are stored as a source of energy for further germination, the fatty acid synthesis pathway is located in the plastids. The first step is the formation of acetyl-ACP (acyl carrier protein) from acetyl-COA and ACP catalyzed by a short chain preferring condensing enzyme, ß-ketoacyl-ACP synthase (KAS) III. Elongation of acetyl-ACP to 16- and 18- carbon fatty acids involves the cyclical action of the following sequence of reactions: condensation with a two-carbon unit from malonyl-ACP to form a longer ß-ketoacyl-ACP (ß-ketoacyl-ACP synthase), reduction of the

keto-function to an alcohol (ß-ketoacyl-ACP reductase), dehydration to form an enoyl-ACP (ß-hydroxyacyl-ACP dehydrase), and finally reduction of the enoyl-ACP to form the elongated saturated acyl-ACP (enoyl-ACP reductase). ß-ketoacyl-ACP synthase I (KAS I), is primarily responsible for elongation up to palmitoyl-ACP (C16:0), whereas ß-ketoacyl-ACP synthase II (KAS II) is predominantly responsible for the final elongation to stearoyl-ACP (C18:0).

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Genes encoding peptide components of ß-ketoacyl-ACP synthases I and II have been cloned from a number of higher plant species, including castor (Ricinus communis) and Brassica species (USPN 5,510,255). KAS I activity was associated with a single synthase protein factor having an approximate molecular weight of 50 kD (synthase factor B) and KAS II activity was associated with a combination of two synthase protein factors, the 50 kD synthase factor B and a 46 kd protein designated synthase factor A. Cloning and sequence of a plant gene encoding a KAS III protein has been reported by Tai and Jaworski (Plant Physiol. (1993) 103:1361-1367).

The end products of plant fatty acid synthetase activities are usually 16- and 18-carbon fatty acids. There are, however, several plant families that store large amounts of 8- to 14-carbon (medium-chain) fatty acids in their oilseeds. Recent studies with *Umbellularia* californica (California bay), a plant that produces seed oil rich in lauric acid (12:0), have demonstrated the existence of a medium-chain-specific isozyme of acyl-ACP thioesterase

in the seed plastids. Subsequent purification of the 12:0-ACP thioesterase from Umbellularia californica led to the cloning of a thioesterase cDNA which was expressed in seeds of Arabidopsis and Brassica resulting in a substantial accumulation of lauric acid in the triglyceride pools of these transgenic seeds (USPN 5,512,482). These results and subsequent studies with medium-chain thioesterases from other plant species have confirmed the chain-length-determining role of acyl-ACP thioesterases during de novo fatty acid biosynthesis (T. Voelker (1996) Genetic Engineering, Ed. J. K. Setlow, Vol. 18, pgs. 111-133).

DESCRIPTION OF THE FIGURES

Figure 1. DNA and translated amino acid sequence of Cuphea

hookeriana KAS factor B clone chKAS B-2 are provided.

Figure 2. DNA and translated amino acid sequence of Cuphea hookeriana KAS factor B clone chKAS B-31-7 are provided.

Figure 3. DNA and translated amino acid sequence of Cuphea hookeriana KAS factor A clone chKAS A-2-7 are provided.

Figure 4. DNA and translated amino acid sequence of Cuphea hookeriana KAS factor A clone chKAS A-1-6 are provided.

Figure 5. DNA and translated amino acid sequence of Cuphea pullcherrima KAS factor B clone cpuKAS B/7-8 are provided.

Figure 6. DNA and translated amino acid sequence of Cuphea pullcherrima KAS factor B clone cpuKAS B/8-7A are provided.

Figure 7. DNA and translated amino acid sequence of Cuphea pullcherrima KAS factor A clone cpuKAS A/p7-6A are provided.

Figure 8. Preliminary DNA sequence of Cuphea pullcherrima KAS factor A clone cpuKAS A/p8-9A is provided.

15 A-2-7 is provided.

- Figure 9. DNA and translated amino acid sequence of Cuphea hookeriana KASIII clone chKASIII-27 are provided.
- Figure 10. The activity profile for purified cpuKAS B/8-7A using various acyl-ACP substrates is provided.
- 5 Figure 11. The activity profile for purified chKAS A-2-7 and chKAS A-1-6 using various acyl-ACP substrates is provided.
 - Figure 12. The activity profile for purified castor KAS factor A using various acyl-ACP substrates is provided.
- 10 Figure 13. The activity profile for purified castor KAS factor B using various acyl-ACP substrates is provided.

 Figure 14. A graph showing the number of plants arranged according to C8:0 content for transgenic plants containing CpFatB1 versus transgenic plants containing CpFatB1 + chKAS
 - Figure 15. Graphs showing the %C10/%C8 ratios in transgenic plants containing ChFatB2 (4804-22-357) and in plants resulting from crosses between 4804-22-357 and 5401-9 (chKAS A-2-7 plants) are provided.
- 20 Figure 16. Graphs showing the %C10 + %C8 contents in transgenic plants containing ChFatB2 (4804-22-357) and in plants resulting from crosses between 4804-22-357 and 5401-9 (chKAS A-2-7 plants) are provided.
- Figure 17. Graphs showing the %C10/%C8 ratios in transgenic plants containing ChFatB2 (4804-22-357) and in plants resulting from crosses between 4804-22-357 and 5413-17 (chKAS A-2-7 + CpFatB1 plants) are provided.
 - Figure 18. Graphs showing the %C10 + %C8 contents in transgenic plants containing ChFatB2 (4804-22-357) and in

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plants resulting from crosses between 4804-22-357 and 5413-17 (chKAS A-2-7 + CpFatB1 plants) are provided.

Figure 19. Graphs showing the %C12:0 in transgenic plants containing Uc FatB1 (LA86DH186) and in plants resulting from crosses with wild type (X WT) and with lines expressing Ch KAS A-2-7.

Figure 20. Graph showing the relative proportions of C12:0 and C14:0 fatty acids in the seeds of transgenic plants containing Uc FatB1 (LA86DH186) and in plants resulting from crosses with wild type (X WT) and with lines expressing Ch KAS A-2-7.

Figure 21. Graphs showing the %C18:0 in transgenic plants containing Garm FatB1 (5266) and in seeds of plants resulting from crosses with wild type (X WT) and with lines expressing Ch KAS A-2-7.

Figure 22. The activity profile of Ch KAS A in protein extracts from transgenic plants containing Ch KAS A-2-7. Extracts were preptreated with the indicated concentrations of cerulenin.

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SUMMARY OF THE INVENTION

By this invention, compositions and methods of use related to ß-ketoacyl-ACP synthase (KAS) are provided. Also of interest are methods and compositions of amino acid and nucleic acid sequences related to biologically active plant synthase(s).

In particular, genes encoding KAS protein factors A and B from Cuphea species are provided. The KAS genes are of interest for use in a variety of applications, and may be

used to provide synthase I and/or synthase II activities in transformed host cells, including bacterial cells, such as E. coli, and plant cells. Synthase activities are distinguished by the preferential activity towards longer and shorter acyl-ACPs as well as by the sensitivity towards the KAS specific inhibitor, cerulenin. Synthase protein preparations having preferential activity towards medium chain length acyl-ACPs are synthase I-type or KAS I. KAS I class is sensitive to inhibition by cerulenin at concentrations as low as 1 µM. Synthases having preferential activity towards longer chain length acyl-ACPs are synthase II-type or KAS II. The KAS enzymes of the II-type are also sensitive to cerulenin, but at higher concentrations (50 μM). Synthase III-type enzymes have preferential activity towards short chain length acyl-ACPs and are insensitive to cerulenin inhibition.

Nucleic acid sequences encoding a synthase protein may be employed in nucleic acid constructs to modulate the amount of synthase activity present in the host cell,

20 especially the relative amounts of synthase I-type, synthase III-type and synthase III-type activity when the host cell is a plant host cell. In addition, nucleic acid constructs may be designed to decrease expression of endogenous synthase in a plant cell as well. One example is the use of an antisense synthase sequence under the control of a promoter capable of expression in at least those plant cells which normally produce the enzyme.

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Of particular interest in the present invention is the coordinate expression of a synthase protein with the

expression of thioesterase proteins. For example, coordinated expression of synthase factor A and a medium-chain thioesterase provides a method for increasing the level of medium-chain fatty acids which may be harvested from transgenic plant seeds. Furthermore, coordinated expression of a synthase factor A gene with plant medium-chain thioesterase proteins also provides a method by which the ratios of various medium-chain fatty acids produced in a transgenic plant may be modified. For example, by expression of a synthase factor A, it is possible to increase the ratio of C10/C8 fatty acids which are produced in plant seed oils as the result of expression of a thioesterase having activity on C8 and C10 fatty acids.

15 DETAILED DESCRIPTION OF THE INVENTION

A plant synthase factor protein of this invention includes a sequence of amino acids or polypeptide which is required for catalyzation of a condensation reaction between an acyl-ACP having a chain length of C2 to C16 and malonyl-ACP in a plant host cell. A particular plant synthase factor protein may be capable of catalyzing a synthase reaction in a plant host cell (for example as a monomer or homodimer) or may be one component of a multiple peptide enzyme which is capable of catalyzing a synthase reaction in a plant host cell, i.e. one peptide of a heterodimer.

Synthase I (KAS I) demonstrates preferential activity towards acyl-ACPs having shorter carbon chains, C2-C14 and is sensitive to inhibition by cerulenin at concentrations of 1µM. Synthase II (KAS II) demonstrates preferential

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activity towards acyl-ACPs having longer carbon chains, C_{14} - C_{16} , and is inhibited by concentrations of cerulenin (50 μ M). Synthase III demonstrates preferential activity towards acyl-CoAs having very short carbon chains, C_{2} to C_{6} , and is insensitive to inhibition by cerulenin.

Synthase factors A and B, and synthase III proteins obtained from medium-chain fatty acid producing plant species of the genus Cuphea are described herein. As described in the following Examples, synthase A from C. hookeriana is naturally expressed at a high level and only in the seeds. C. hookeriana synthase B is expressed at low levels in all tissues examined. Expression of synthase A and synthase B factors in E. coli and purification of the resulting proteins is employed to determine activity of the 15 various synthase factors. Results of these analyses indicate that synthase factor A from Cuphea hookeriana has the greatest activity on 6:0-ACP substrates, whereas synthase factor B from Cuphea pullcherrima has greatest activity on 14:0-ACP. Similar studies with synthase factors A and B from castor demonstrate similar activity profiles 20 between the factor B synthase proteins from Cuphea and castor. The synthase A clone from castor, however, demonstrates a preference for 14:0-ACP substrate.

Expression of a Cuphea hookeriana KAS A protein in transgenic plant seeds which normally do not produce medium-chain fatty acids does not result in any detectable modification of the fatty acid types and contents produced in such seeds. However, when Cuphea hookeriana KAS A protein is expressed in conjunction with expression of a

medium-chain acyl-ACP thioesterase capable of providing for production of C8 and C10 fatty acids in plant seed oils, increases in the levels of medium-chain fatty acids over the levels obtainable by expression of the medium-chain thioesterase alone are observed. In addition, where significant amounts of C8 and C10 fatty acids are produced as the result of medium-chain thioesterase expression, co-expression of a Cuphea KAS A protein also results in an alteration of the proportion of the C8 and C10 fatty acids that are obtained. For example, an increased proportion of C10 fatty acids may be obtained by co-expression of Cuphea hookeriana ChFatB2 thioesterase and a chKAS A synthase factor proteins.

Furthermore, when Cuphea hookeriana KAS A protein is expressed in conjunction with expression of a medium-chain acyl-ACP thioesterase capable of providing for production of C12 fatty acids in plant seed oils, increases in the levels of medium-chain fatty acids over the levels obtainable by expression of the medium-chain thioesterase alone are also observed. In addition, where significant amounts of C12 and C14 fatty acids are produced as the result of medium-chain thioesterase expression, co-expression of a Cuphea KAS A protein also results in an alteration of the proportion of the C12 and C14 fatty acids that are obtained. For example, an increased proportion of C12 fatty acids may be obtained by co-expression of Uc FatB1 thioesterase and a chKAS A synthase factor proteins.

However, when Cuphea hookeriana KAS A protein is expressed in conjunction with the expression of a long-chain

acyl-ACP thioesterase capable of providing for production of C18 and C18:1 fatty acids in plant seed oils, no effect on the production of long chain fatty acids was observed. Furthermore, when plants transformed to express a long chain acyl-ACP thioesterase from mangosteen (GarmFatA1, U.S. Patent Application No. 08/440,845), which preferentially hydrolyzes C18:0 and C18:1 fatty acyl-ACPs, are crossed with nontransformed control plants, a significant reduction in the levels of C18:0 is obtained. Similar reductions are also observed in the levels of C18:0 in the seeds of plants resulting from crosses between plants transformed to express the GarmFatA1 and plants expressing the Cuphea hookeriana KAS A protein.

Thus, the instant invention provides methods of increasing and/or altering the medium-chain fatty acid compositions in transgenic plant seed oils by co-expression of medium-chain acyl-ACP thioesterases with synthase factor proteins. Furthermore, various combinations of synthase factors and medium-chain thioesterases may be achieved depending upon the particular fatty acids desired. For example, for increased production of C14 fatty acids, synthase protein factors may be expressed in combination with a C14 thioesterase, for example from Cuphea palustris or nutmeg may be employed (WO 96/23892). In addition, thioesterase expression may be combined with a number of different synthase factor proteins for additional effects on medium-chain fatty acid composition.

Synthases of use in the present invention include modified amino acid sequences, such as sequences which have

been mutated, truncated, increased and the like, as well as such sequences which are partially or wholly artificially synthesized. The synthase protein encoding sequences provided herein may be employed in probes for further screening or used in genetic engineering constructs for transcription or transcription and translation in host cells, especially plant host cells. One skilled in the art will readily recognize that antibody preparations, nucleic acid probes (DNA and RNA) and the like may be prepared and used to screen and recover synthases and/or synthase nucleic acid sequences from other sources. Typically, a homologously related nucleic acid sequence will show at least about 60% homology, and more preferably at least about 70% homology, between the R. communis synthase and the given plant synthase of interest, excluding any deletions which may be present. Homology is determined upon comparison of sequence information, nucleic acid or amino acid, or through hybridization reactions.

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Recombinant constructs containing a nucleic acid sequence encoding a synthase protein factor or nucleic acid sequences encoding a synthase protein factor and a medium-chain acyl-ACP thioesterase may be prepared by methods well known in the art. Constructs may be designed to produce synthase in either prokaryotic or eukaryotic cells. The increased expression of a synthase in a plant cell, particularly in conjunction with expression of medium-chain thioesterases, or decreasing the amount of endogenous synthase observed in plant cells are of special interest.

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Synthase protein factors may be used, alone or in combination, to catalyze the elongating condensation reactions of fatty acid synthesis depending upon the desired result. For example, rate influencing synthase activity may reside in synthase I-type, synthase III-type, synthase III-type or in a combination of these enzymes. Furthermore, synthase activities may rely on a combination of the various synthase factors described herein.

Constructs which contain elements to provide the 10 transcription and translation of a nucleic acid sequence of interest in a host cell are "expression cassettes". Depending upon the host, the regulatory regions will vary, including regions from structural genes from viruses, plasmid or chromosomal genes, or the like. For expression in prokaryotic or eukaryotic microorganisms, particularly unicellular hosts, a wide variety of constitutive or regulatable promoters may be employed. Among transcriptional initiation regions which have been described are regions from bacterial and yeast hosts, such as E. coli, 20 B. subtilis, Saccharomyces cerevisiae, including genes such as ß-galactosidase, T7 polymerase, trp-lac (tac), trp E and the like.

An expression cassette for expression of synthase in a plant cell will include, in the 5' to 3' direction of transcription, a transcription and translation initiation control regulatory region (also known as a "promoter") functional in a plant cell, a nucleic acid sequence encoding a synthase, and a transcription termination region.

Numerous transcription initiation regions are available

which provide for a wide variety of constitutive or regulatable, e.g., inducible, transcription of the desaturase structural gene. Among transcriptional initiation regions used for plants are such regions

5 associated with cauliflower mosaic viruses (35S, 19S), and structural genes such as for nopaline synthase or mannopine synthase or napin and ACP promoters, etc. The transcription/ translation initiation regions corresponding to such structural genes are found immediately 5' upstream to the respective start codons. Thus, depending upon the intended use, different promoters may be desired.

Of special interest in this invention are the use of promoters which are capable of preferentially expressing the synthase in seed tissue, in particular, at early stages of seed oil formation. Examples of such seed-specific promoters include the region immediately 5' upstream of a napin or seed ACP genes such as described in USPN 5,420,034, desaturase genes such as described in Thompson et al (Proc. Nat. Acad. Sci. (1991) 88:2578-2582), or a Bce-4 gene such as described in USPN 5,530,194. Alternatively, the use of the 5' regulatory region associated with the plant synthase structural gene, i.e., the region immediately 5' upstream to a plant synthase structural gene and/or the transcription termination regions found immediately 3' downstream to the plant synthase structural gene, may often be desired. general, promoters will be selected based upon their expression profile which may change given the particular application.

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In addition, one may choose to provide for the transcription or transcription and translation of one or more other sequences of interest in concert with the expression or anti-sense of the synthase sequence,

5 particularly medium-chain plant thioesterases such as described in USPN 5,512,482, to affect alterations in the amounts and/or composition of plant oils.

When one wishes to provide a plant transformed for the combined effect of more than one nucleic acid sequence of interest, a separate nucleic acid construct may be provided for each or the constructs may both be present on the same plant transformation construct. The constructs may be introduced into the host cells by the same or different methods, including the introduction of such a trait by crossing transgenic plants via traditional plant breeding methods, so long as the resulting product is a plant having both characteristics integrated into its genome.

Normally, included with the DNA construct will be a structural gene having the necessary regulatory regions for expression in a host and providing for selection of transformed cells. The gene may provide for resistance to a cytotoxic agent, e.g. antibiotic, heavy metal, toxin, etc., complementation providing prototrophy to an auxotrophic host, viral immunity or the like. Depending upon the number of different host species into which the expression construct or components thereof are introduced, one or more markers may be employed, where different conditions for selection are used for the different hosts.

The manner in which the DNA construct is introduced into the plant host is not critical to this invention. Any method which provides for efficient transformation may be employed. Various methods for plant cell transformation include the use of Ti- or Ri-plasmids, microinjection, electroporation, liposome fusion, DNA bombardment or the like. In many instances, it will be desirable to have the construct bordered on one or both sides by T-DNA, particularly having the left and right borders, more particularly the right border. This is particularly useful when the construct uses A. tumefaciens or A. rhizogenes as a mode for transformation, although the T-DNA borders may find use with other modes of transformation.

The expression constructs may be employed with a wide variety of plant life, particularly plant life involved in the production of vegetable oils. These plants include, but are not limited to rapeseed, peanut, sunflower, safflower, cotton, soybean, corn and oilseed palm.

For transformation of plant cells using Agrobacterium, explants may be combined and incubated with the transformed Agrobacterium for sufficient time for transformation, the bacteria killed, and the plant cells cultured in an appropriate selective medium. Once callus forms, shoot formation can be encouraged by employing the appropriate plant hormones in accordance with known methods and the shoots transferred to rooting medium for regeneration of plants. The plants may then be grown to seed and the seed used to establish repetitive generations and for isolation of vegetable oils.

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The invention now being generally described, it will be more readily understood by reference to the following examples which are included for purposes of illustration only and are not intended to limit the present invention.

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EXAMPLES

Example 1 Cuphea KAS Factor A and B Gene Cloning

Total RNA isolated from developing seeds of Cuphea hookeriana and Cuphea pullcherrima was used for cDNA synthesis in commercial 1-based cloning vectors. For cloning each type of KAS gene, approximately 400,000-500,000 unamplified recombinant phage were plated and the plaques transferred to nitrocellulose. For KAS factor B cloning from C. hookeriana, a mixed probe containing Brassica napus KAS factor B and Ricinus communis (Castor) KAS factor B radiolabeled cDNA's was used. Similarly, a mixed probe containing Brassica napus KAS factor A and Ricinus communis KAS factor A cDNA clones was used to obtain C. hookeriana 20 KAS factor A genes. For KASIII, a spinach KASIII cDNA clone obtained from Dr. Jan Jaworski was radiolabeled and used as a probe to isolate a KASIII clone from C. hookeriana. For KAS B and KAS A cloning from C. pullcherrima, C. hookeriana KAS B and KAS A genes chKAS B-2 and chKAS A-2-7 (see below) were radiolabeled and used as probes.

DNA sequence and translated amino acid sequence for Cuphea KAS clones are provided in Figures 1-9. Cuphea hookeriana KAS factor B clones chKAS B-2 and chKAS B-31-7

are provided in Figures 1 and 2. Neither of the clones is full length. Cuphea hookeriana KAS Factor A clones chKAS A-2-7 and chKAS A-1-6 are provided in Figures 3 and 4. chKAS A-2-7 contains the entire encoding sequence for the KAS factor protein. Based on comparison with other plant synthase proteins, the transit peptide is believed to be represented in the amino acids encoded by nucleotides 125-466. chKAS A-1-6 is not a full length clone although some transit peptide encoding sequence is present. Nucleotides 1-180 represent transit peptide encoding sequence, and the mature protein encoding sequence is believed to begin at nucleotide 181.

Cuphea pullcherrima KAS factor B clones cpuKAS B/7-8 and cpuKAS B/8-7A are provided in Figures 5 and 6. Both of the clones contain the entire encoding sequences for the KAS factor B proteins. The first 35 amino acids of cpuKAS B/7-8 are believed to represent the transit peptide, with the mature protein encoding sequence beginning at nucleotide 233. The first 39 amino acids of cpuKAS B/8-7A are believed to represent the transit peptide, with the mature protein encoding sequence beginning at nucleotide 209. Cuphea pullcherrima KAS factor A clones cpuKAS A/p7-6A and cpuKAS A-p8-9A are provided in Figures 7 and 8. Both of the clones contain the entire encoding sequences for the KAS factor A proteins. Translated amino acid sequence of cpuKAS A/p7-6A is provided. The mature protein is believed to begin at the lysine residue encoded 595-597, and the first 126 amino acids are believed to represent the transit peptide. DNA sequence of KAS A clone cpuKAS A-p8-9A is preliminary.

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Further analysis will be conducted to determine final DNA sequence and reveal the amino acid sequence encoded by this gene.

DNA and translated amino acid sequence of Cuphea hookeriana KASIII clone chKASIII-27 is provided in Figure 9. The encoding sequence from nucleotides 37-144 of chKASIII-27 are believed to encode a transit peptide, and the presumed mature protein encoding sequence is from nucleotides 145-1233.

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Deduced amino acid sequence of the C. hookeriana KAS factor B and KAS factor A cDNA's reveals strong homology to the Brassica napus and Ricinus communis clones previously reported. The C. hookeriana KAS factor B clone is more homologous to the Ricinus and Brassica KAS factor B clones (94% and 91% respectively) than it is to the Ricinus and Brassica KAS factor A clones (60% for both). Furthermore, the C. hookeriana KAS factor A clone is more homologous to the Ricinus and Brassica KAS factor A clones (85% and 82% respectively) than it is the Ricinus and Brassica KAS factor B clone (60% for both). The C. hookeriana KAS factor B cDNAs designated as chKAS B-2 and chKAS B-31-7 are 96% identical within the mature portion of the polypeptide. Similarly, the deduced amino acid sequence of the mature protein regions of the C. hookeriana KAS factor A clones chKAS A-2-7 and chKAS A-1-6 are 96% identical. The C. pullcherrima KAS clones also demonstrate homology to the R. communis and Brassica napus KAS clones. The mature protein portion of all of the KAS factor A family members in the different Cuphea species are 95% identical. Similarly the

mature protein portion of the KAS factor B genes in Cuphea are also 95-97% identical with each other. However there is only approximately 60% sequence identity between KAS factor B and KAS factor A clones either within the same or different species of Cuphea.

Example 2 Levels and Patterns of Expression

To examine tissue specificity of KAS expression in Cuphea hookeriana, Northern blot analysis was conducted 10 using total RNA isolated from seed, root, leaf and flower tissue. Two separate but identical blots were hybridized with either chKAS B-31-7 or chKAS A-2-7 coding region probes. The data from this RNA blot analysis indicate that KAS B is expressed at a similar level in all tissues 15 examined, whereas KAS A expression is detected only in the seed. These results also demonstrate a different level of expression for each of the synthases. KAS A is an abundant message, whereas KAS B is expressed at low levels. Furthermore, even under highly stringent hybridization conditions (65_C, 0.1 X SSC, 0.5% SDS), the KAS A probe hybridizes equally well with two seed transcripts of 2.3 and 1.9 kb. The larger hybridizing band is likely the transcript of the KAS A-2-7 gene since the size of its cDNA is 2046bp, and the number of clones obtained from cDNA 25 screening corresponds well with the apparent mobility of the mRNA and its abundance on the blot.

Example 3 Expression of Plant KAS Genes in E.coli

DNA fragments encoding the mature polypeptide of the Cuphea hookeriana KAS A cDNAs and the Cuphea pullcherrima

5 KAS B cDNAs were obtained by PCR and cloned into a QIAexpress expression vector (Qiagene). Experimental conditions for maximum level of expression were determined for all of these clones and the parameters for highest level of soluble fraction were identified. Cells are grown in ECLB media containing 1M sorbitol and 2.5 mM betaine overnight and subcultured as a 1:4 dilution in the same medium. Cells are then grown for 2 hours (to approximately .6-.8 O.D.) and induced with 0.4 mM IPTG and allowed to grow for 5 more hours.

Enzyme activity of the affinity purified recombinant enzymes obtained from over-expression of the chKAS A-2-7 and cpuKAS B/8-7A clones was measured using a wide range of acyl-ACP substrates (6:0- to 16:1-ACP). The activity profile for cpuKAS B/8-7A is provided in Fig.10. The data demonstrate that the enzyme is active with all acyl-ACP substrates examined, although activity on 6:0 to 14:0-ACP substrates is substantially greater than the activity on 16:0 and 16:1 substrates.

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The activity profile of the *C. hookeriana* KAS A clones chKAS A-2-7 and chKAS A-1-6 is provided in Figure 11. The *C. hookeriana* KAS A clones are most active with C:6, and have the least activity with C:16:0 substrates. However, the activity of this clone on even the preferred C6:0 substrate

is 50 fold lower than the activity of the *C. pullcherrima* KAS B clones.

A fragment containing the mature protein encoding portion of a R. communis KAS factor A clone was also cloned into a QIAexpress expression vector, expressed in E. coli and the enzyme affinity purified as described above. The activity profile for castor KAS A is provided in Figure 12. Highest activity is observed with C14:0 substrates, although some activity is also seen with C6:0 and C16:1. In comparison, the activity profile obtained from purified R. communis KAS factor B also using the QIAexpress expression system is provided in Figure 13. The KAS B clone demonstrates substantially higher levels of activity (10 fold and higher) than the R. communis KAS A clone. The preference of the KAS factor B for 6:0- to 14:0-ACP substrates is consistent with the previous observations that this protein provides KAS I activity.

Example 4 KAS and TE Expression in Transgenic Seed

Dehesh et al. (1996) Plant Physiol. 110:203-210) and the chKAS A-2-7 were PCR amplified, sequenced, and cloned into a napin expression cassette. The napin/cp FatB1 and the napin/KAS A-2-7 fusions were ligated separately into the binary vector pCGN1558 (McBride and Summerfelt (Pl.Mol.Biol. (1990) 14:269-276) and transformed into A. tumefaciens, EHA101. The resulting CpFatB1 binary construct is pCGN5400 and the chKAS A-2-7 construct is pCGN5401. Agrobacterium mediated transformation of a Brassica napus canola variety

15

was carried out as described by Radke et al. (Theor. Appl. Genet. (1988) 75:685-694; Plant Cell Reports (1992) 11:499-505). Several transgenic events were produced for each of the pCGN5400 and pCGN5401 constructs.

A double gene construct containing a napin/cpFatB1 expression construct in combination with a napin/chKAS A-2-7 expression construct was also assembled, ligated into a binary vector and used for co-cultivation of a canola Brassica variety. The binary construct containing the chFatB1 and chKAS A-2-7 expression constructs is pCGN5413.

Fatty acid analysis of 26 transgenic lines containing chKAS A-2-7 (5401 lines) showed no significant changes in the oil content or profile as compared to similar analyses of wild type canola seeds of the transformed variety.

Fatty acid analysis of 36 transgenic lines containing cpFatB1 (5400 lines) showed increased levels of C:8 and C:10 in transgenic seeds. The highest level of C:8 observed in a pool seed sample was 4.2 mol%. The C:10 levels were between 30 and 35% of the C:8 content. Fatty acid analysis of 25 transgenic lines containing the TE/KAS A tandem (5413 lines) demonstrated an overall increase in both C:8 and C:10 levels relative to those observed with TE containing lines (5400) alone. In lines containing the cpFatB1 construct alone, the average level of C:8 average were 1.5 mol%, whereas the C:8 average levels in TE/KAS A tandem containing lines was 2.37 mol%. The ratio of C:8 to C:10 remained constant in both populations. The number of transgenic events relative to the C:8 content are presented in Figure 14. These data show that the transgenic events with tandem TE/KAS A construct

yield more lines with higher levels of C:8 than those events with single TE construct. For example, several lines containing nearly 7 mole% C8 were obtained with the TE/KAS A pCGN5413 construct, whereas the highest C8 containing line from the pCGN5400 TE alone transformation contained 4.2 mole% C8.

Half seed analysis of the T3 generation of transgenic canola plants expressing a ChFatB2 (*C. hookeriana* thioesterase; Dehesh et al. (1996) The Plant Journal 9:167-172) indicate that these plant can accumulate up to 22 weight% (33 mol%) of 8:0 and 10:0 fatty acids (4804-22-357). Segregation analysis shows that these transformants contain two loci and that they are now homozygous. Selected plants grown from these half seeds were transferred into the greenhouse and later crossed with T1 transformants that had been transformed with either Cuphea hookeriana KAS A (5401) alone or KAS A/CpFatB1 double constructs (5413).

Fatty acid analysis of several events resulting from the crosses between transgenic lines containing ChFatB2 (4804-22-357) and chKAS A-2-7 (5401-9), reveal an increase in the ratio of C:10/C:8 levels (Figure 15). This C:10/C:8 ratio in nearly all of the transgenic events containing ChFatB2 TE alone fluctuates between 3 and 6, whereas in the F1 generation of transgenic containing both the TE and the KAS A-2-7, the ratio can be as high as 22. This increase in C:10 levels is accompanied by an increase in the total C:8 and C:10 content (Figure 16). The sum of the C:8 and C:10 fatty acids in the heterozygous F1 lines is as high as those in the homozygous parent line (4804-22-357), whereas the

20

heterozygous lines usually contain substantially less C:8 and C:10 than the homozygous lines.

Similar results were observed in F1 generation seeds resulting from crosses performed between 4804-22-357 (ChFatB2) and the 5413-17 event (CpFatB1 and chKAS A-2-7 tandem). Levels of C:8 and C:10 in the 5413-17 line were 6.3 and 2.8 mol% respectively. Data presented in Figure 17 show that there is shift towards C:10 fatty acids as was observed with the 4804-22-357 (ChFatB2) x 5401-9 (chKAS A-2-7) crosses. Furthermore, Figure 18 indicates the presence of two separate populations of heterozygotes. containing approximately 9-11 weight percent C:10 + C:8 are believed to represent offspring containing a single copy of the ChFatB1 TE gene and no copies of the CpFatB1 and chKAS A genes from 5413. Those plants containing approximately 15-20 weight percent C:10 + C:8 are believed to represent the heterozygotes containing a single ChFatB1 TE gene as well as the CpFatBl and chKAS A genes from 5413. Thus, the level of the C:10 + C:8 fatty acids does not decrease to 50% of that detected in parent lines when a copy of the ChKAS A gene is present.

To further characterize the chain length specificity of the Cuphea hookeriana KAS A enzyme, crosses between transgenic Brassica napus lines containing a California Bay (Umbellularia californica) 12:0 specific thioesterase, Uc FatB1 (USPN 5,344,771) and chKAS A-2-7 (5401-9) were made. Half seed analysis of transgenic plants containing Uc fatB1 have previuosly indicated that these plants can accumulate up to 52 mol% C12:0 in the seed oil of homozygous dihaploid

lines (LA86DH186). Crosses between the line LA86DH186 and untransformed control *Brassica* demonstrated a decrease in the C12:0 levels.

However, crosses between LA86DH186 and the 5401-9

hemizygous line led to an accumulation of up to 57 mol%

C12:0 in the seed oil of F1 progeny (Figure 19).

Interestingly, in crosses with LA86DH186 x untransformed control line and LA86DH186 x 5401-9, levels of C14:0 in the seeds of the F1 progeny decreased to 50% of the levels

obtained in homozygous LA86DH186 lines (Figure 20).

Furthermore, increases in the proportion of C12:0 fatty acid resulted in a substantial decline in the proportions of all the long-chain fatty acyl groups (C16:0, C18:0, C18:2, and C18:3). These results indicate that the ChKAS A-2-7 is an enzyme with substrate specificity ranging from C6:0 to C10:0-ACP, and that its over-expression ultimately reduces the longer chain acyl-ACP pools.

Further evidence is obtained in support of the chain length specificity of the ChKAS A-2-7 in crosses of the 5401-9 line with a transgenic line (5266) expressing an 18:1/18:0 TE from Garcinia mangostana (GarmFatAl, US patent application No. 08/440,845). Transgenic Brassica line 5266 has been shown to accumulate up to 24 mol% C18:0 in the seed oil of homozygous lines (Figure 21). However, in the seed oil of F1 progeny of crosses between 5266 and 5401-9 levels of C18:0 were reduced to approximately 12 mol%. Furthermore, levels of C16:0 generated from these crosses was similar to the levels obtained from the seed oil of nontransgenic control plants.

Example 5 In vitro Analysis of Plant KAS Enzymes

Seed extracts were prepared from developing seeds of nontransgenic controls or transgenic Brassica expressing chKAS A-2-7 as described in Slabaugh et al. (Plant Journal, 5 1998 in press) and Leonard et al. (Plant Journal, 1998, in press). In vitro fatty acid synthesis assays were performed as described by Post-Beittenmiller (J. Biol. Chem. (1991), 266:1858-1865). Extracts were concentrated by ammonium sulfate precipitation and desalting using P-6 columns (Bio-Rad, Hercules, CA). Reactions (65µl) contained 0.1M 10 Tris/HCl (pH 8.0), 1 mM dithiothreitol, 25 mM recombinant spinach ACP1, 1 mM NADH, 2 mM NADPH, 50 µM malonyl-CoA, 10 μM [1-14C]acetyl-CoA (50 mCi/mmol), 1mg/ml BSA, and 0.25 mg/ml seed protein. Selected seed extracts were preincubated with cerulenin at 23°C for 10 min. Reaction products were separated on an 18% acrlamide gel containing 2.25M urea, electroblotted onto to nitrocellulose and quntitated by phosporimaging using Image QuaNT software (Molecular Dynamics, Sunnyvale, CA). Authentic acyl-ACPs were run in parallel, immunoblotted and finally detected by anti-ACP serum to confirm fatty acid chain lengths.

The results (Figure 22) indicate that the fatty acid synthesis capabilities of transgenic Brasica (5401-9) seed extracts was greater than that obtained from in the nontransgenic controls as measured by the relative abundance of C8:0- and C10:0-ACP at all time points tested. In addition, pretreatment of the extracts with cerulenin, markedly reduced the synthesis of longer chain fatty acids in both the transgenic and nontransgenic control seed

extracts. However, the extension of the spinach-ACP was much less inhibited in the seed extracts from the transgenic lines than in the seed extracts of nontransgenic control Brassica.

These data further support that Ch KAS A-2-7 is a condensing enzyme active on medium chain acyl-ACPs, and that expression of this enzyme in plants results in enlarged substrate pools to be hydrolyzed by medium-chain specific thioesterases. Furthermore, these data suggest that chKAS A-2-7 also is a cerulenin-resistant condensing enzyme.

All publications and patent applications mentioned in this specification are indicative of the level of skill of those skilled in the art to which this invention pertains.

- All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.
- Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious that certain changes and modifications may be practiced within the scope of the appended claim.

WO 98/46776

PCT/US98/07114

28

MISSING UPON TIME OF PUBLICATION

- 13: The construct of Claim 5 wherein said encoding sequence is cpuKAS A/p8-9A.
- 14. The construct of Claim 5 wherein said encoding sequence is chKASIII-27.
- 15. An improved method for producing medium-chain fatty acids in transgenic plant seeds by expression of a plant medium-chain thioesterase protein heterologous to said transgenic plant,

the improvement comprising expression of a plant synthase
factor protein heterologous to said transgenic plant in
conjunction with expression of said plant medium-chain
thioesterase, whereby the percentage of medium-chain fatty
acids produced in seeds expressing both a plant synthase factor
protein and a plant medium-chain thioesterase protein is
increased as compared to the percentage of medium-chain fatty
acids produced in seeds expressing only said plant medium-chain

- 16. The method of Claim 15 wherein said medium-chain thioesterase protein is a ChFatB2 protein.
- 20 17. The method of Claim 15 wherein said medium-chain thioesterase protein is a CpFatBl protein.

thioesterase protein.

- 18. The method of Claim 15 wherein said medium-chain thioesterase protein is a C12 preferring thioesterase from California bay.
- 19. The method of Claim 15 wherein said plant synthase factor protein is expressed from a construct according to Claim 1.
 - 20. The method of Claim 19 wherein said synthase factor A protein is from a Cuphea species.

21. The method of Claim 20 wherein said Cuphea species is C. hookeriana or C. pullcherrima.

22. A method of altering the medium-chain fatty acid composition in plant seeds expressing a heterologous plant medium-chain preferring thioesterase, wherein said method comprises

providing for expression of a plant synthase factor protein heterologous to said transgenic plant in conjunction with expression of a plant medium-chain thioesterase protein heterologous to said transgenic plant, whereby the composition of medium-chain fatty acids produced in said seeds is modified as compared to the composition of medium-chain fatty acids produced in seeds expressing said plant medium-chain thioesterase protein in the absence of expression of said plant synthase factor protein.

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- 23. The method of Claim 22 wherein said medium-chain thioesterase protein is a ChFatB2 protein.
- 24. The method of Claim 22 wherein said medium-chain thioesterase protein is a CpFatBl protein.
- 25. The method of Claim 22 wherein said medium-chain thioesterase protein is a C12 preferring thioesterase from California bay.
- 26. The method of Claim 22 wherein said plant synthase factor protein is expressed from a construct according to Claim 25 1.
 - 27. The method of Claim 26 wherein said synthase factor A protein is from a Cuphea species.
 - 28. The method of Claim 27 wherein said Cuphea species is C. hookeriana or C. pullcherrima.

WO 98/46776 PCT/US98/07114

- 29. The method of Claim 22 wherein said fatty acid composition is enriched for C10 fatty acids.
- 30. The method of Claim 22 wherein said fatty acid composition is enriched for C12 fatty acids.
- 31. The method of Claim 22 wherein said fatty acid composition is enriched for at least one medium chain fatty acid and at least one other medium chain fatty acid is decreased.
- 32. The method of Claim 31 wherein said enriched fatty 10 acid is C12 and said decreased fatty acid is C14.

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48	96	144	192	240	. 288	336	384
	•						
66C 61y	AAG Lys	GGT Gly	CAC	GGG Gly	TCA	GCT Ala	ACT Thr
CCG	TCC	GGT	GGT G1y	ATG Met	TAT	GCC Ala	GGC Z
CCC	CTC	ATG Met	AAG Lys	AAC Asn	AAC Asn	GCT	GGA
gat Asp	CGC Arg	GGA Gly	GAG	ACA Thr	CCA	CAT	GCT Ala
GTG Val	GAC Asp	ACA Thr	ATC Ile	ATT Ile	66C 61y	TTC Phe	ATT
CTA Leu	GCC Ala	GGA Gly	CTT	GCC	ATG Met	TGC Cys	ATG Met
GAA Glu	GGT Gly	GTC Val	TCT Ser	TAT Tyr	CTC	TAC Tyr	CTT Leu
CTA Leu	CTC	CTG	CAG Gln	CCC	GGT Gly	AAC Asn	gat Asp
GCT Ala	GAT Asp	GTG Val	GTT Val	ATC Ile	TTT Phe	TCC Ser	GCT
GCC	GCC Ala	GGA Gly	GGG G1y	TTC	GAA Glu	ACT	GAG Glu
GCG Ala	CGA Arg	GCC Ala	GAC	TTC Phe	ATC Ile	GCC Ala	GGT G1y
GTG Val	GCA Ala	AGA Arg	TCT Ser	CCT	GCT	TGT Cys	CGT Arg
GCG	TCG Ser	GAG Glu	TTC Phe	ACC Thr	CTC	GCA Ala	CGC Arg
ACC Thr	AAT Asn	AAG Lys	GTC Val	ATC Ile	CTG	ACT Thr	ATC
TCC	AGG Arg	GAC Asp	ACT Thr	AAA Lys	GCC Ala	TCC Ser	CAT His
AGC	TGC Cys	ATC Ile	CTG	CGG Arg	TCT	ATT Ile	AAT Asn

FIGURE 1 1 OF 4

432	480	528	576	624	672	720	768
AGG Arg	TGG Trp	TTG	ATT Ile	ACT	AGC	GCT	ATC
TGC Cys	CCC	GTG Val	ATT Ile	ATG Met	AGT Ser	AAT Asn	GCC Ala
GCT	AGG Arg	GGA Gly	CCG	CAC His	GAG Glu	ATA Ile	AAT Asn
GTG Val	TCT Ser	GCT Ala	GCA Ala	TAT Tyr	ATT Ile	\mathtt{TAC}	ATA Ile
TTT Phe	GCC	GGT Gly	66A 61y	GCT	TGC Cys	AAT Asn	GAG Glu
66C G1Y	ACT	GAA Glu	CGA Arg	GAT Asp	TCT Ser	GTC Val	GCC
GGA Gly	CAG Gln	GGT Gly	aga Arg	TGT Cys	TCT Ser	GAG Glu	SAT CTC ASP Leu FIGURE 1 2 OF 4
TTG Leu	CCG	ATG Met	ATG Met	AAC Asn	GTC Val	GAA Glu	GAT ASP FIGU
666 61y	GAC Asp	GTG Val	GCA Ala	ATC Ile	GGT Gly	CCT	GGG G1y
ATT Ile	GAT Asp	TTT Phe	CAT His	GCA Ala	CTT	TCA Ser	GCT Ala
CCA Pro	AAC Asn	GGT Gly	GAA Glu	GGT Gly	GGT Gly	GTC Val	CTA
ATT Ile	AGG Arg	GAT Asp	TTG	GGA Gly	GAT Asp	GGC Gly	ACT
ATC Ile	CAA Gln	CGT Arg	AGC Ser	TTG Leu	GCT Ala	GCT Ala	TCT Ser
GCA Ala	TCT Ser	GAC Asp	GAG Glu	TAT Tyr	AGG Arg	GAT Asp	ACT Thr
GCC	TTG Leu	AAA Lys	ATG Met	GAG Glu	CCA Pro	GAA Glu	GCG Ala
GAG Glu	GCT	GAT ASD	GTG Val	GCA	GAT Asp	CTT Leu	CAT His

816	864	912	096	1008	1056	1116	
AAG AAG GTT TTC AAG AAC ACA AAG GAT ATC AAA ATT AAT GCA ACT AAG	TCA ATG ATC GGA CAC TGT CTT GGA GCA TCT GGA GGT CTT GAA GCT ATA	GCG ACT ATT AAG GGA ATA AAC ACC GGC TGG CTT CAT CCC AGC ATT AAT	CAA TTC AAT CCT GAG CCA TCG GTG GAG TTC GAC ACT GTT GCC AAC AAG	AAG CAG CAA CAC GAA GTT AAC GTT GCG ATC TCG AAT TCA TTC GGA TTT	GGA GGC CAC AAC TCA GTC GTG GCT TTC TCG GCT TTC AAG CCA TGATTA	CCCATTICAC AAGGIACTIG TCATIGAGAA TACGGAITAT GGACTIGCAG AGTAATITCC CCATGITIGI CGGAAGAGCA TATTACCACG GTIGICCGIC AAACCCATIT AGGATACIGI	FIGURE 1
Lys Lys Val Phe Lys Asn Thr Lys Asp Ile Lys Ile Asn Ala Thr Lys	Ser Met Ile Gly His Cys Leu Gly Ala Ser Gly Gly Leu Glu Ala Ile	Ala Thr Ile Lys Gly Ile Asn Thr Gly Trp Leu His Pro Ser Ile Asn	Gln Phe Asn Pro Glu Pro Ser Val Glu Phe Asp Thr Val Ala Asn Lys	Lys Gln Gln His Glu Val Asn Val Ala Ile Ser Asn Ser Phe Gly Phe	Gly Gly His Asn Ser Val Val Ala Phe Ser Ala Phe Lys Pro		3 OF 4

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FIGURE	4 OF 4

*	TCTCAAAAA	GGAAGTGCCG TCTCAAAAA	ACTITIGITI GIATIGGAAA GGAAGIGCCG ICICAAAAAA AAAAAAAAA AA

Sequence Range: 1 to 1704

40 0											
GTG Val		GCA Ala>		TCT Ser>	06	GAC Asp>	240	cGG Arg>	CTC Leu>		GAA Glu>
GNG		TCG	140	GAC	19	ATC Ile		ATC Ile	AGG Arg		CTC
ACC	90	AAT Asn	, ,	GTC		TTA		CAG Gln	NO AGG Arg	330	GCT_CTC Ala_Leu
30 TCC Ser		AGG Arg		GAC Asp		AGC	230	GGC Gly	280 GAC AC ASP AI		AAG Lys
AGC		TGC	0	TCC	180	ATC	N	GGC Gly	AAC Asn		AAG Lys
TGG	80	GGC	13(GGC Gly		GGG Gly		TTC	AAG Lys	320	GGG
20 AGC Ser		CCG Pro		TTC		AGC	0	AGG Arg	270 GGG Gly	m	GCC
AAA Lys		CCC		GTA Val	170	GAG Glu	220	ACC	GAC Asp		GTC
AAC Asn	7.0	GAT Asp	120	TCC	П	GGC Gly		CCC	ATC Ile	0	TGC ATT Cys Ile
10 AAA GGG Lys Gly	•	GTG Val		GTC		TCC Ser		TTC Phe	260 TAC TYE	310	TGC
AAA Lys		CTA		CTC	160	CTC	210	AAG Lys	2 GGA Gly		TAC
ACT		GAA Glu	110	${\tt GGC}$	16	CTC		TCC Ser	ACG		CGC
CTC	¢09	CTA		ATG GGC (Met Gly 1		AAG Lys		GCT Ala	250 AAC GCG Asn Ala	300	CTC
ACC		GCT		GGC Gly		GAA Glu	200	GAC	25 AAC Asn		TGC
TTA		GCC	100	GCC Ala	150	TAC Tyr	(4)	TTC Phe	TTC		GAT Asp
AAA Lys	20	GCG Ala	1(CGA Arg		TAT Tyr		CGC Arg	GGA Gly	90	GAC

FIGURE 2 1/5

	AGA Arg>	430	TCT Ser>	480	CCG Pro>	GCC Ala>		TGT Cys>		CGA Arg>	670	ATT Ile>	
380	GAG Glu	43	TTC Phe		TCC	CTT Leu		GCA Ala	620	CGC Arg	6	ATC Ile	
m	AAG Lys		GTC Val		ATC Ile	CTG	570	ACT Thr	w	ATC Ile		GCA Ala	
	GAT		ACC Thr	470	AAG Lys	520 GCT CTA Ala Lei		TCA		CAT His		GCT	
0	ATT Ile	420	CTA Leu	4	CGG Arg	TCT Ser		ATT Ile	610	AAT Asn	099 *	GAG Glu	
370	AAG Lys		66C 61y		CAC His	GGG G1y	260	TCG	61	GCC Ala		ACT Thr	
	TCC		$_{\rm GGT}$	460	GGT Gly	510 ATG Met	u,	TAT Tyr		GCT		GGA Gly	
	CTC	410	ATG Met	46	AAA Lys	AAC Asn		AAC		GCC	650	GGA Gly	M 10
360	AGC	7	GGT Gly		GAG Glu	ACA Thr	550	CCA	009	\mathtt{TAT}		GCT	FIGURE 2/5
	GAA Glu		ACT		ATC Ile	500 ATT Ile	Ŋ	GGC Gly		TTT Phe		ATT Ile	Ħ
	GGT	400	GGA Gly	450	CTC	GCC		ATG		TGC Cys	640	ATG	
350	GGC	4(GTT Val		AAT Asn	TAT Tyr		CTG	290	TAC	9	CTC	
(*)	CTC		CTA		CAG Gln	490 T CCC e Pro	540	ggr Gly		AAC Asn		GAC	
	GAT Asp		GTG Val	440	GTT Val	AT		TTG		TCC		GCT	
340	TCC Ser	390	GGA Gly	•	666 G1y	TTC		GAT Asp	580	ACT Thr	630	GAG Glu	
34	AAT Asn		GCT		GAC Asp	TTT Phe	30	ATC Ile	ũ	GCT		GGC G1y	

SUBSTITUTE SHEET (RULE 26)

720	AGG Arg>	GAT Asp>		TTG Leu>		GGA Gly>		GAT Asp>	096	GGG Gly>	ACT Thr>
7.5							910		9		
	CAA Gln	CGT Arg		AGC	860	TTG	o)	GCT		GCT Ala	TCC
	TCT	60 GAC ASP	810	GAG Glu	w	TAT Tyr		AGG Arg		GAT ASP	O ACT Thr
710	TTA Leu	7. AAG Lys		ATG Met		GAA Glu		CCA	950	GAA Glu	1000 T GCG AC' S Ala Th
•	GCT	GAT Asp		GTT Val	850	GCA Ala	900	GAT Asp	01	CTG	C.A.
	AGG Arg	TGG Trp	800	$_{ m Leu}$	80	ATT Ile		ACT Thr		AGT	GCT Ala
700	TGC	750 CCG Pro	ω,	GTA Val		ATT Ile		ATG Met	940	AGC	990 AAT Asn
7(GCC	AGG Arg		GGA G1y		CCG	890	CAT His	6	GAG Glu	ATA Ile
	GTT Val	TCA	0	GCT	840	GCG Ala	ω	TAT		ATT Ile	TAC
	TTC Phe	740 GCC Ala	790	$\begin{array}{c} GGG\\ G1Y \end{array}$		GGA Gly		GCT		TGC	980 AAT Asn
069	GGA Gly	ACT Thr		GAA Glu		CGA Arg	0	GAT	930	TCT Ser	g GTC Val
	GGA Gly	CAG Gln		GGC Gly	830	AAA Lys	88	TGT		TCC	GAG Glu
	TTA Leu	30 CCT Pro	780	ATG Met	w	ATG Met		AAT		GTC Val	70 GAA Glu
089	GGG G1y	730 GAC CC ASP Px		GTG Val		GCA		GTC Val	920	GGT Gly	970 CCT GA Pro Gl
•	ATT Ile	gat Asp		TTT Phe	820	CAT His	870	GCA Ala	01	CTT	TCA
	CCA	AAT Asn	70	GGT Gly	8	GAA Glu		GGT Gly		$^{\rm GGG}_{\rm G1Y}$	GTC Val

FIGURE 2 3/5

	AAG Lys>		CAC His>	0	GGA Gly>	1200	GAG Glu>	GAA Glu>		TCA Ser>		GCA
	TTC Phe	1100	GGA Gly	1150	AAG Lys	-	CCC	CAT His		AAC Asn	1340	AAT
1050	GTT Val	11	ATC Ile		ATT Ile		AAT Asn	CAA Gln	1290	CAC His	ਜ	GGT TCA AAT
Н	AAG		ATG		ACA Thr	1190	TTC Phe	1240 CAG CAA Gln Gln		GGC Gly		GGT
	AAG Lys	0	TCG	1140	GCG Ala	11	CAA Gln	AAG Lys		GGA Gly	30	CTC
1040	GCC ATC Ala Ile	1090	AAG Lys	**	ATT Ile		AAC Asn	AAG Lys	1280	TTC	1330	TTA
10	GCC		ACT		GCC Ala	000	ATA Ile	1230 GCC AAC Ala Asn	11	GGA Gly		TGA
	AAT Asn		GCA	1130	CTT GAA GCC Leu Glu Ala	1180	AGC			TTC		CCA
0	ATA Ile	1080	AAT	11			CCC	GTT Val	0.2	TCA	1320	AAG Lys
1030	GAG	Н	ATC		GGT Gly		CAT	1220 GAC ACA ASP Thr	1270	AAT		TTC
	GCC		ACA Thr	02	GGG Gly	1170	CTT	12 GAC ASP		TCA		GCC
	CTT	1070	ATC Ile	1120	TCA	•	TGG Trp	TTC		ATC Ile	1310	TCA
1020	GAT	10	GAA Glu		GCA Ala		GGC Gly	1210 GTG GAA Val Glu	1260	GCT Ala	Н	TTC
-	GGG G1Y		AAG Lys		GGA Gly	1160	ACC			GTT Val		GCT
	GCT	0.9	ACC Thr	1110	CTT	H	ACC	TCA		AAT Asn	00	GTA Val
10	CTT	1060	AAC		TGT Cys		ATA Ile	CCA Pro	20	GTG Val	1300	GTT Val

FIGURE 2

FIGURE 2 5/5

AATTIGITGC IGAGACAGIG AGCITCAACT IGCAGAGCAA ITITITACAI GCCIIGICGI CGGAAGAGCG TAATACCGGG ATAGTTCCTT GATAGTTCAT TTAGGATGTT TTACTGCAAT AATCGAAGAT TATTTCCATT CTAATCCAGT CTCCGNCGAG TTTGAGAATC TATCTGTTTG CCCTTGTCAA TGGCATTTAA GATAAGCTTA TAAAAAAAA AAAAAAAAA AAAACTCGAG GGGGGCCCG GTACCCAATT CGCCCTATAG TGAGTCGTAT GACAATTCAC TGTCCGTCGG TATTAGAAAG AACGAGGCAA GATTTTGTTT CATGTTTGTG TTTGTATTAC TTTCTTTTTG

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w	TGG?	13	GGTC	TGG Trp		TCC Ser		TCC		TGC	360	GGA Gly
	CTAG		GCTCAGGTGT	ACG		CGT Arg	0	CTC	310	CCT	.,	TTC
50	3A A(110		TGT Cys	210	CCA	260	ACT Thr		GAT		CIC
	rcta(સં	3GCT(160 TTC Phe	7	GAC		AGG		CTC	0	
	CCGCTCTAGA ACTAGTGGAT		GGTCGGCTCA	CCT		AAC		CGG	300	TGC	350	GCT TCC Ala Ser
40		100		TCC	0	GAC	250	CGC	М	CAA		TTC
	gcggrggcgg		TTCTTACTTG	150 Grr GCG Val Ala	200	TCC		CGT		TTC		GGA Gly
	gcg			Grr Val		TCA		TCC	0	ACC	340	AAC Asn
30	CACC	90	AGTT	ATG		ACT	240	CTC	290	TCC		GAT Asp
	ACTAAAGGGA ACAAAAGCTG GAGCTCCACC		GGCACGAGTT	140 rcr rgc ser Cys	190	CCC	73	CGC		GGA Gly		GGG G1y
20	G GA	80		14 TCT Ser		ATG		CTC		CGC Arg	330	CTC
73	AGCT	80	AATT	GCT		TGC Cys	0	CGG Arg	280	CTC	(-)	TTC
	CAAA		CCCCCGGGCT GCAGGAATTC	ACC Thr	180	GCA Ala	230	AAG Lys		TCC		CGC
10	GA A	70	CT G	130 GCG	Н	GCT		CAC His		TGC	320	CAA
	AAGG		5552	ATG Met		GTA		TCC Ser	270	CAT His	35	CAG Gln
	ACTA		CCCC	TCCA	170	CTC	220	CTT Leu	(1)	TCC		AAC

FIGURE 3 1/6

ACT		GAA Glu		GTG Val		TAC	009	AAC	TCT Ser		GAC
CGC		CAG	00	GTT Val	550	GTT Val	9	GAG AAC Glu Asn	AAG Lys		ATG
66C G1y	450	GCA	2(GTA Val		GAT Asp		ATA Ile	ATC	069	AGG
400 CTC Leu		CCT		CGA Arg		CCC	290	GAG Glu	640 GAG Glu	v	GAG Glu
AGG		CAA		AGG	540	GAC	55	AGT	GGA Gly		TCC
CTG	440	ATG Met	490	CAA Gln	٠,	CAT		ATA Ile	GCC	680	TTC
390 CAC	4	GCT		AAG Lys		GGC Gly		GGC Gly	630 ATT Ile	39	AAG Lys
660 61y		GTG Val		ACC	30	CTA	580	AGT	AGA Arg		CCA
CGC		GCT	480	GCT	53	CCT		ATA Ile	ACG Thr		GCC
380 TCA AAȚ Ser Asn	430	ATG Met	,	CCT		ACT		GGA Gly	620 TTT CCC Phe Pro	670	GTG Val
38 TCA Ser		GTC		AAA Lys		GTG Val	570	GAC	62 TTT Phe		TGG Trp
CGT		GAG	470	AAG Lys	520	GTG Val	٥,	CTA	CAG Gln		GGC Gly
CTT Leu	420	GGG G1y	4	AAT Asn		66C G 1y		CTC	TCT Ser	099	ტ∢
370 CCT Pro	,	TCC		ACA Thr		ATG	260	AAT Asn	610 TGC Cys	v	ACA Thr
AAG Lys		CAT His		TCC	510	GGT Gly	26	AAC	GAC Asp		TCC
TCC	410	TCC	460	GTC	u ,	ACA Thr	-	TAC	TTC Phe	650	TTT Phe

FIGURE 3 2 OF 6

	GAT Asp		TGT Cys	840	GAT Asp	TGT Cys		GAC		ACA		GAA Glu
740	GCA Ala	790	AAG Lys	ω	AGC	TTT Phe		ATG Met	980	GCA Ala	1030	GGC
	TTA		AGA Arg		TTC Phe	CCC	930	GCA Ala	96	TGT Cys	, ,	AAA Lys
	GCA		AAA Lys	0	GTA Val	880 AGT Ser	01	CTT Leu		GCC		ATC Ile
730	AAA Lys	780	AAT Asn	830	AAG Lys	ATC		ATT Ile		ACT Thr	1020	CAC ATA His Ile
7	AAG	7	CIC		ATG Met	AAG Lys	920	GCT Ala	970	TCA	1)	CAC His
	GGC		GAG Glu		GGT Gly	870 AAG Lys	92	TCC		ATA Ile		AAC
0	GCA	0	AAA Lys	820	GGC Gly	8 TAT TYE		GGA Gly		TCG	10	GCG
720	ACT	770	ATG		TTG	TCA		ATG Met	096	TAT	1010	GCT Ala
	CTG		GCG Ala		GGA Gly	860 AGG ACT TO Arg Thr So	910	AAT Asn	O,	AAC Asn		AAT Asn
	* ATG Met		GAT	810	TCC	86 AGG Arg		ACA		CCT		CTG
710	TAC	760	GAA Glu	ω	66c G1y	CTG		ACC Thr	950	GGC Gly	1000	ATA Ile
	CIT		ACT		ATT Ile	GCT	006	TTT TCT Phe Ser	9,	ATG		TGT
	ATG Met		ATC Ile	0	CTC	850 GAA Glu	0,			TGG Trp		TTC
700	TTC	750	GGA Gly	800	GTT Val	ATT Ile		CCT		GGA Gly	066	AAC Asn
7	AAG Lys	7	GGT Gly		GGA Gly	TCC	890	GTA Val	940	TTG	3,	AGT

FIGURE 3 3 OF 6

1080	GTT Val	AAT Asn		TTT Phe		CAT		AGT Ser	1320	GCT Ala	TCG
ñ	CCT	AAT Asn		GGA Gly	0	GAG Glu	1270	GGG G1y	13	GGA	GTC Val
	TTA	AGG Arg	1170	GAT Asp	122(TTA Leu	11	GGT Gly		GAA Glu	GGA
7.0	GTT Val	1120 CAG Gln	H	CGT		GAG Glu		CTA	07	CCT	1360 CAG TCC GGA Gln Ser Gly
1070	GCC	TCA		AAT Asn		GAG Glu	1260	TTT Phe	1310	CAC His	CAG Gln
	GCG	TTG	9	AGT	1210	CTT Leu	1	GAA Glu		CCT	GCT
	TCG GAT Ser Asp	1110 CGA GCT Arg Ala	1160	GAC	• •	CTT Leu		GCG		GAG Glu	1350 GCC TTG Ala Leu
1060	TCG			TGG		TTA	0.0	TAT Tyr	1300	ACC	13 GCC Ala
	GGC G1y	TGC		CCA	1200	GTT Val	125	ATT	ν,	ATG Met	aag Lys
	GGT Gly	1100 GTA GCA Val Ala	1150	AGA Arg	H	GGA Gly		ACC		CAC His	1340 ATA GAG Ile Glu
1050	TGT			TCG		GCT		GCA Ala	1290	TAC	1340 ATA G2 Ile G
Н	CTT	TTC		GCT	06	GGA Gly	1240	GGT Gly	Ħ	GCC	TGC Cys
	ATG	GGT Gly	1140	AAA Lys	1190	GAA Glu		AGA Arg		GAC Asp	CTC
1040	* ATG	1090 GGA G1y	-	ACC		GGA Gly		AAA Lys	80	TGC Cys	1330 GTG ATC Val Ile
10	GAC	TTG Leu		CCT		ATG Met	1230	AAG Lys	1280	ACT	
	GCA Ala	GGT G1Y	1130	GAC	1180	GTG Val	H	GCA		TTC Phe	GGT Gly

FIGURE 3 4 OF 6

	GCT Ala		AAC		CTT	1560	AGG	GGC Gly		GTC Val		TCC
	CCT	0	CAA	1510	CTT Leu	15	ATA Ile	GAA Glu		AAG Lys	00	AAC TCA Asn Ser
1410	ACT Thr	1460	GGC G1y	П	CAC His		GCA	GAC Asp	1650	CTG Leu	1700	
14	TCC Ser		TTC Phe		$_{\rm G1y}^{\rm GGT}$	0.9	CAG	1600 CCG	'n	AAA Lys		CAT His
	ACT Thr		TGT	1500	ATC Ile	1550	GTT Val	GAC		GAG Glu		GGC Gly
0	GCA Ala	1450	CAC His	15			GTA Val	GAA Glu	40	AAG Lys	1690	GGC
1400	CAT	П	GCC Ala		TCG		GCA Ala	1590 AAT TTG Asn Leu	1640	AAG		TTC
	GCG Ala			0.0	ACC AAA TCG Thr Lys Ser	1540	GTT Val			CCT		666 G1y
	AAT Asn	1440	CAA GCT CTC Gln Ala Leu	1490		-	GCA Ala	ATT		GGC Gly	1680	TCA TTT Ser Phe
1390	ATA Ile	14	CAA		TCC		GAA Glu	1580 CCA AAT Pro Asn	1630	CTC GTC Leu Val	ન	TCA
7	TAC		TAC Tyr		AAT Asn	1530	GTA Val					AAT Asn
	AAT Asn	0.0	GAA Glu	1480	GTG Val	ä	GGT.GGC Gly Gly	ATC.CAT Ile His		CTG	1670	TTG TCC Leu Ser
1380	GTA Val	1430	AAG Lys	,,,	AGA Arg				1620	AAA Lys	16	
13	GAC Asp		ATC Ile		CTG	50	GCT	1570 TGG Trp	त्न	GCA Ala		GGT Gly
	GAA		GAT	1470	GAG	1520	GGA Gly	1 GGA Gly		GAT Asp		GTC
1370	AGG	1420	GGA G	1,	AGT		GGA G1y	ACA	1610	GTG Val	1660	AAG Lys

FIGURE 3 5 OF 6

OF 6	
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1760	CCC TGC AAC TAG A AAAGAGTCTG TGGAAGCCGA GAGTCTTTGA Pro Cys Asn ***	1820	GGCTACTCGA	1880	TTGTCCCTTT	1940	CTTTTCGAAT	2000	ATATTTGAA		
1750	TGGAAGCCGA	1810	GAGATAGACC	1870	AGATCACTGC	1930		1990	TTTGTAATGC		
1740	A AAAGAGTCTG	1800	CTCTGAAACC GAGATAGACC GGCTACTCGA	1860	GGGGATGCCA AAGATACTCC TTGCCGGTAT TGGTGTTAAG AGATCACTGC TTGTCCCTTT	1920	TATTTTCTTC TTCTTTGAG AGCTTTAACC GAGGTAGTCG TATTTTCGAG	1980	TATCGGATCA ATGTGTTTCT TCTAAGATCA TTTGTAATGC ATATTTTGAA	2040	AAACCACATC TCAGTATGCA AAATAAAAA AAAAAAAAA
1730	GC AAC TAG /	1790	GAACTCATGC ACGTTAGTAG CTTCTTATGC	1850	TTGCCGGTAT	1910	AGCTTTAACC	1970	ATGTGTTTCT	2030	АДАДАДАД
1720	GCC Ala	1780	ACGTTAGTAG	1840	AAGATACTCC	1900	TTCTTTTGAG	1960	TATCGGATCA	2020	TCAGTATGCA
1710	ATA CTA TTT Ile Leu Phe	1770	GAACTCATGC	1830	GGGGATGCCA	1890	TATTTTCTTC	1950	ACATGTTCGT	2010	AAACCACATC

GCATCCTTGT TCGGATCCAG GCCCATCCGC ACCACCCGCA GGCACCGGAG GCTCAATCGA GCTTCCCCTT CCGGGGAGGC AATGGCTGTG GCTCTGCAAC CTGCACAGGA AGTTACCACA CGGCACGAGG TCACCTCTTA CCTCGCCTGC TTCGAGCCCT GCCATGACTA CTACACCTCC GCT CAA TTT CCT ACG AGA ATT GCT GGA GAG ATC AAG TCT TTC TCC ACA Ala Gln Phe Pro Thr Arg Ile Ala Gly Glu Ile Lys Ser Phe Ser Thr> CTG CTT GAT GGA ACG AGT GGC ATA AGT GAG ATA GAG ACC TTT GAT TGT Leu Leu Asp Gly Thr Ser Gly Ile Ser Glu Ile Glu Thr Phe Asp Cys> AAG AAG AAG CCA AGT ATC AAA CAG CGG CGA GTA GTT GTG ACT GGA ATG Lys Lys Lys Pro Ser Ile Lys Gln Arg Arg Val Val Val Thr Gly Met> GGT GTG GTG ACT CCT CTA GGC CAT GAC CCT GAT GTT TTC TAC AAT AAT GIY Val Val Thr Pro Leu Gly His Asp Pro Asp Val Phe Tyr Asn Asn> Sequence Range: 1 to 1921

FIGURE 4

420	ATG Met>	ATC Ile>		CTC Leu>		GAA Glu>	610	TTC Phe>	099	TGG Trp>	TTT Phe>
	TTC	GGA G1y		GTT Val	260	ATT	61	CCT		GGA Gly	AAC Asn
	GAC AAG ASP LYS	460 T GGT n Gly	510	GGA Gly	σ,	GCC		GTA Val		TTG Leu	700 ACG AGT Thr Ser
410	GAC	AA As		AAA TGC Lys'Cys		GAT Asp		TGT	029	GAC TTG Asp Leu	
•	ATG Met	ACA Thr			550	AAT Asn	009	TTT Phe	v	ATG	GCA
	AGG Arg	TTA Leu	200	AGA Arg	5.	TTC		CCC		GCA Ala	TGT Cys
400	AAG Lys	450 GCA Ala	۵,	AAA Lys		GTA Val		AAT Asn	640	CTT Leu	690 GCT Ala
4	TCC	AAA Lys		GAT Asp		AAG Lys	290	ATG	9	ATG	ACT Thr
	CTC	AAG Lys	490	CTA	540	ATG Met	υ,	AAG Lys		GCT Ala	TCT Ser
	AAG Lys	440 GGC Gly	4	GAG Glu		GGA Gly		AAG Lys		TCA	80 ATA Ile
390	CCG	GCC		AAA Lys		$_{\rm GGT}^{\rm GGT}$	580	\mathtt{TAT}	630	GGA Gly	TCG
	GCC Ala	ACT		ATG Met	530	ATG Met	28	TCA		ATG Met	TAC
	GTG Val	430 G CTG	480	GTG Val	,	GCA		ATT Ile		AAT Asn	670 CCC AAC TA Pro Asn Ty
380	TGG Trp	43 ATG Met		GAT Asp		TCA	,	AGG Arg	620	ACA Thr	67 CCC Pro
• •	GGT Gly	TAC		GAA Glu	520	GGC Gly	570	CTA Leu	W .	ACC Thr	GGC Gly
	GAT Asp	CTT Leu	470	ACC Thr	52	ATT Ile		GCC Ala		GCT	ATG

FIGURE 4 2/6

	GTG Val>		GGA Gly>	0	ACT Thr>	006	666 G1y>	AAA Lys>		TGC Cys>		ATT Ile>
	GAT Asp	800	ATG Met	850	CCT		ATG Met	AAG Lys		ACT Thr	1040	GTG Val
750	GCA Ala	ω	GGT Gly		GAC		GTT Val	940 T GCA S Ala	990	TTC Phe	ਜ ਜ	GGA Gly
	GAA Glu		ATT Ile		GCC Ala	068	TTT Phe	E.		AGT		GCT Ala
	AGA GGC Arg Gly	790	CCT Pro	840	AAT Asn	w	GGA Gly	GAG Glu		GGA Gly	30	GAT GGA ASP Gly
740	AGA Arg	75	ATA Ile		AGA Arg		GAT Asp	TTA	980	GGT Gly	1030	GAT Asp
-	ATC Ile		ATC Ile		CAG Gln	880	CGT Arg	930 GAG Glu	σ,	CTA		CCT
	ATA Ile		GTA Val	830	TCA	80	AAT Asn	GAG Glu		TTT Phe		CAC His
730	CAC	780	GCG	w	TTG		AGT	CTA	970	GAA Glu	1020	CCT
73	AAC		GAT Asp		GCT		GAC	920 CTA Leu	, O	GCA Ala		GAG Glu
	GCG		TCA	820	CGA Arg	870	TGG Trp	CTA		TAC		ACC
	GCT	170	GGC Gly	8	TGC		CCA	GTG Val		ATT Ile	1010	CAC ATG His Met
720	AAT Asn		$^{\rm GGG}_{\rm G1Y}$		GCA		AGA Arg	910 GCT GGA Ala Gly	960	ACT Thr	Ä	
	CTG		TGC		GTT Val	860	TCA			GCG Ala		TAC
	ATC Ile	760	CTT Leu	810	TTT Phe	~	GCT Ala	GGA Gly		GGT Gly	00	GCC
710	TGT Cys	76	ATG Met		GGT Gly		AAA Lys	GAA Glu	950	AGA Arg	1000	GAT ASD

FIGURE 4

	•							•	•			
06	GAA GAC Glu Asp>	1140	ATC Ile>	TTA Leu>		GCC Ala>		TGG Trp>		ACC Thr>	1380	GGT Gly>
1090			GAT Asp	GAG Glu		GCA Ala	1280	666 61y	1330	GAT Asp		GTC Val
	AGG Arg		GGA Gly	30 AAC Asn	1230	GGA Gly	12	ACT Thr	٠	GTG Val		AAG Lys
	TCT	1130	GCT	1180 AAC AAC Asn Asn	П	CTC		AGG Arg		GGC Gly	1370	ATT Ile
1080	GTC Val	H	CCA	caa Gin		CTT	0,	GCA ATA Ala Ile	1320	GAA Glu	13	AAC Asn
••	GGA Gly		ACT Thr	GGC Gly	1220	CAC His	1270	GCA		GAT Asp		CTG
	TCA	20	TCC	1170 TGT TTC (13	GGT Gly		CAG Gln		CCA	20	AGA Arg
1070	CAG Gln	1120	ACA Thr	TGT		ATT Ile		GTT Val	1310	AAC Asn	1360	GAG Glu
H	GCT		GCC	CAC His	10	ATG Met	1260	GTA Val	ਜ	GAA Glu		AAG Lys
	TTG		CAT His	1160 TT ATC	1210	TCA	.,	TCA		TTG		AAG Lys
1060	GCT	1110	GCA Ala	ខ្ម		AAA Lys		GTT Val	00	ATT AAT Ile Asn	1350	CCT
10	AAG Lys		AAT Asn	GCT		ACC Thr	1250	GCA Ala	1300	ATT Ile	•	GGC Gly
	GAG Glu		ATA Ile	1150 TAC CAA Tyr Gln	1200	TCT	Ä	GAA Glu		AAT Asn		GTG Val
	TGC ATA C	1100	TAC			AAT		GTG Val		CCG Pro	1340	CTC
1050			AAT Asn	GAG Glu		GTG Val	40	GGT Gly	1290	CAT His	H	TTG
	CTC		GTA Val	AAA Lys	1190	AAA Lys	1240	GGT Gly	•	ATC Ile		AAA Lys

FIGURE 4

CTC TTC	1480	ITCAAA	1540	CATGCCCATG	1600	GGCGACACAG	1660	TTTCTGAAAT	1720	AGTCAGTGAA GAAGAGAACA	1780	TTTATCGCCG	1840	TITIGIGGGI TAAAATITGI AAAACTAGAC GACTGGITIG TITICICTIG AICAIIGGAG
1420 TCG TCC ATA Ser Ser Ile	1470	CATGTGTGGA ATTCTACTCA ATCTATCAAA	1530	AGCATGTTGG TAGCTCCTTA CGTCTCTAGA CATGCCCATG	1590	AGTTTTGTGT CGGGAGCTGT AGTCGGAACC ATGACGGATT GAGTACTCAT GGCGACACAG	1650	TCCCATTTT TTTCTGAAAT	1710	AGTCAGTGAA	1770	CCCTTTGTTT TGCTCTCTAT TTTATCGCCG	1830	TTTTCTCTTG
GGG TTT GGT GGG CAC AAC TCG TCC GIY Phe Gly Gly His Asn Ser Ser	1460	GTGGA ATTCI	1520	TAGCTCCTTA	1580	ATGACGGATT	1640	CTATTCATTA	1700	CGTTTCATCG	1760		1820	GACTGGTTTG
1400 GGG TTT GGT G Gly Phe Gly G	1450		1510	AGCATGTTGG	1570	AGTCGGAACC	1630	TTGCTAGAAT TGTTAGAGCA	1690	TACTTTCGAG	1750	GGGCACGTAG TAACCATTTG	1810	AAAACTAGAC
TCA TTC Ser Phe	1440	AAC TAG GGCGTTT ASn ***>	1500	TGAGGACTCC	1560	CGGGAGCTGT	1620		1680	CTCCCTCCTT ACGGTAGTTG TACTTTCGAG	1740		1800	TAAAATTTGT
1390 TTG TCT AAT Leu Ser Asn	1430	GCC CCT TAC Ala Pro Tyr	1490	GCTGAAGTTT	1550	AGTTTTGTGT	1610	GATATACTCC	1670	CTCCCTCCTT	1730	AAGCTAACTC	1790	TTTTGTGGGT

FIGURE 4 5/6

1900

1890

1880

1870

1860

1850

1910 1920 * AAAAAAAA AAAAAAAA A

ATGTATGGCC ATATTTGCCT TTCATTGATG ATAAAAAAA AAAAAAAA AAAAAAAA

FIGURE 4 6/6

	ATG GGA Met Gly 110 CGC TAC AFG TYF	
CTGGTACGCC TGCAGGTACC GGTCCGGAAT TCCCGGGTCG ACCCACGCGT CGGTCTTCCC ACTCCGATCC TTGGCTTCTC CGCCATCTTC CGCCATCTTC TTGGCTTTCT CGCCATCCTC CGCCGCT CGCCATCCTC CGCCATC CTC CGC CTC CT	ACC AGG TTC GGC GGC CAG ATT CGT GGC Thr Arg Phe Gly Gly Gln Ile Arg Gly 105 GAC GGC AAA AAC GAC AGG CGG CTT GAT ASP Gly Lys Asn Asp Arg Arg Leu Asp Alg 115	

FIGURE 5

553	601	649	697	745	793	841	688
GCC	666 G1y	CTT Leu	GCC Ala 190	ATG Met	TGC Cys	ATG Met	66C 61y
GGT Gly	GTT Val	TCT	${\tt TAT} \\ {\tt TYT}$	CTG Leu 205	TAC Tyr	CTT	GGA Gly
CTC Leu 140	CTG	CAA Gln	CCC	GGT Gly	AAC Asn 220	GAT Asp	TTG Leu
GAT Asp	GTG Val 155	GTT Val	ATC Ile	CTC	TCC	GCT Ala 235	GGG Gly
GCC Ala	GGA G1y	GGG G1y 170	TTC Phe	GAA Glu	ACT Thr	GAG Glu	ATT 11e 250
gac Asp	GCC	GAC ASP	TTC Phe 185	ATT Ile	GCC Ala	GGT Gly	CCA
GAG Glu	AGA Arg	TCT	CCT	GCT Ala 200	TGT	CGT Arg	ATT Ile
CTT Leu 135	GAG Glu	TTC Phe	ACC Thr	CIC	GCA Ala 215	CGC Arg	ATC Ile
TCT Ser	AAG Lys 150	GTC Val	ATC Ile	CTG	ACT Thr	ATC Ile 230	GCA Ala
AAG Lys	GAC Asp	ACT Thr 165	AAA Lys	GCC Ala	TCC	CAT His	GCC Ala 245
AAG Lys	ATC Ile	CTG	CGG Arg 180	TCT	ATT Ile	AAT Asn	GAG Glu
666 617	AAG Lys	GGT	CAC His	GGG Gly 195	TCA	GCT Ala	ACT Thr
GCC Ala 130	TCC	GGT Gly	GGT G1y	ATG Met	TAT Tyr 210	GCT	66C 61y
GTC Val	CTC Leu 145	ATG Met	AAG Lys	AAC Asn	AAC Asn	GCT Ala 225	GGA Gly
ATT Ile	CGC Arg	GGA G1Y 160	GAG Glu	ACA Thr	CCA	CAT His	GCT Ala 240
TGC Cys	GAC	ACA Thr	ATC 11e 175	ATT Ile	66C 61y	TTC	ATT Ile

FIGURE 5

937	985	1033	1081	1129	1176	1224	1272
ACT Thr 270	GAA Glu	CGA Arg	GAT Asp	TCT	GTC Val 350	GCC	AAA Lys
CAG Gln	GGT G1y 285	AAA Lys	TGT Cys	TCC	GAG Glu	CTC Leu 365	ATC Ile
CCT Pro	ATG Met	ATG Met 300	AAC Asn	GTC Val	GAA Glu	gat Asp	GAT ASP 380
GAC Asp	GTG Val	GCA Ala	ATC Ile 315	GGT Gly	CCT	666 G1y	AAG Lys
gat Asp	TTT Phe	CAT His	GCA Ala	CTC Leu 330	TCA	GCT Ala	ACA
AAC Asn 265	GGT Gly	GAA Glu	GGT Gly	GGT Gly	GTC Val 345	CTA	AAC
AGG Arg	GAT Asp 280	TTG Leu	GGA Gly	GAT Asp	GGC	ACT Thr 360	AAG Lys
CAA Gln	CGT	AGC Ser 295	TTG Leu	GCT Ala	GCT Ala	TCT Ser	TTC Phe 375
TCT Ser	GAC Asp	GAG Glu	TAT Tyr 310	AGG Arg	GAT Asp	ACT	GTT Val
CTG Leu	AAA Lys	CTG	GAG Glu	CCA Pro 325	GAA Glu	GCG Ala	AAG Lys
GCT Ala 260	GAT	GTG Val	GCA	GAC Asp	CTT Leu 340	CAT His	AAG Lys
AGG Arg	TGG Trp 275	TTG Leu	ATT Ile	ACT	AGC	GCT Ala 355	ATC
TGC	CCC	GTG Val 290	ATT Ile	ATG Met	AGT Ser	AAT Asn	GCC Ala 370
GCT Ala	AGG Arg	GGA Gly	CCT Pro 305	CAC His	GAG Glu	ATA Ile	AAT Asn
GrG Val	TCT Ser	GCT Ala	GCA Ala	TAT Tyr 320	ATT Ile	TAC	ATA Ile
TTT Phe 255	GCC Ala	GGT	GGA Gly	GCT	TGC Cys 335	AAT Asn	GAG Glu

FIGURE 5

1320	1368	1416	1464	1512	1569	1629	1689	1712
ATT AAT GCA ACT AAG TCA ATG ATC GGA CAC TGT CTT GGA GCC TCT GGA Ile Asn Ala Thr Lys Ser Met Ile Gly His Cys Leu Gly Ala Ser Gly 385	GGT CTT GAA GCT ATA GCG ACT ATT AAG GGA ATA AAC ACC GGC TGG CTT Gly Leu Glu Ala Ile Ala Thr Ile Lys Gly Ile Asn Thr Gly Trp Leu 400	CAT CCC AGC ATT AAT CAA TTC AAT CCT GAG CCA TCC GTG GAG TTC GAC His Pro Ser Ile Asn Gln Phe Asn Pro Glu Pro Ser Val Glu Phe Asp 415	ACT GTT GCC AAC AAG CAG CAA CAC GAA GTT AAT GTT GCG ATC TCG Thr Val Ala Asn Lys Lys Gln Gln His Glu Val Asn Val Ala Ile Ser 435	AAT TCA TTT GGA TTC GGA GGC CAC AAC TCA GTC GTG GCT TTC TCG GCT Asn Ser Phe Gly Phe Gly Gly His Asn Ser Val Val Ala Phe Ser Ala 450	TTC AAG CCA TGA TTACC CATTTCACAA GGCACTTGTC ATTGAGAGTA CGGTTGTTCG Phe Lys Pro 465	TCAAACCCAT TTAGGATACT GTTCTATGTA AAAAAAGTA AGGATTATCA CTTTCCCTTC	TAATCCTGTC TCCAGTTTGA GAATGAAATT ATATTTATTT TAAAAAAAAA	GGCCGCTCTA GAGGATCCAA GCT

FIGURE 5

Sequence Range: 1 to 1802

					•							
	09 *	TTATCTCCGC	.0 CCT TCC Pro Ser		TCC Ser	210	CGT Arg		CGG Arg		GTC Val	CTA
		TATC	110 rcc ccr ser Pro	160	TCC	7	ATC Ile		AAG Lys		GAC	350 ATC AGC Ile Ser
	20		CAC T His S		CCC		GTC Val		AAG Lys	300	TCC	35 ATC Ile
		GCCT	CTC C Leu H		TCC	0	CCC	250	CCC	(*)	GGC	GGC
		GGTCGACCCA CGCGTCCGGG CTTTCCGACC ACATTTCATT TCTTGCCTCG	100 TCC C Ser L	150	AAT Asn	200	CIC		GAC		TTC	AGC Ser
	40	ATT	CAA T	Н	CTC		AGC		TCC	290	GTC Val	340 GAG Glu
		TTTC			CGC		cgc gcc Arg Ala	240	GAG Glu	23	TCC	66C G1y
		ACA	υ «Σ	0	TTC	190	CGC	73			GTC Val	TCC
	30	GACC	70 80 90 GGCTCCTCCG CCGTCGTTCG CCGCCGCCGC ATG	140	CCC		CGT		GCC CCC AAG Ala Pro Lys		CTC	330 CTG CTC Leu Leu
		TTCC	9009		GAG Glu		CTC	0	CCC	280	GGC	
	20	G CI	80 CG CC		CTC	180	CCC	230	GCC		ATG	AAG Lys
	7	9900	8 GTTC	130	CCT	П	CGC Arg		TCC		ACC GGC Thr Gly	320 TAC GAC Tyr Asp
7		GCGT	CGTC		TCC		CTC		GCC	270	ACC Thr	
to 1802	10	CA	70 CG C		CCC Pro	0	GCT Ala	220	ACC	(4	ATC Ile	TAC
1 to		GACC	CCTC	120	cGC Arg	170	GCC		GCC		GTC Val	GCC
Range:		GGTC	CGCT	Н	CTC		GCC		GCT	260	GTC Val	310 GAC ASP
4)												

FIGURE 6 1/5

	CAG Gln	450	CGG Arg		GCT Ala		AAG Lys	GTC Val		ATC Ile	069	CTG	•
400	GGC Gly	4.	GAC Asp		AAG Lys		GAT Asp	O ACT Thr	640	AAG Lys	•	GCG	
	GCC		AAC Asn		AAG Lys	540	ATT Ile	590 CTA A(Leu T)		CGG Arg		TCT Ser	
	TTC	0	AAG Lys	490	GGC Gly	ហ	AAG Lys	66C 61y		CAC	0	GGG Gly	
390	AGG Arg	440	GGC Gly		GCC		TCC	GGT Gly	. 089	$_{\rm GLY}^{\rm GGT}$	680	ATG Met	
m	ACC Thr		GAC		GTC	0	CTC	580 ATG Met	Ψ	AAA Lys		AAC Asn	
	CCC		ATC Ile	480	TGC ATT Cys Ile	53	TCC	GGT Gly		GAG Glu		ACA Thr	9
380	TTC	430	TAC	4	TGC		CAA Gln.	ACC Thr	620	ATC Ile	670	ATT Ile	URE
38	AAA Lys		66C G1y		TAC Tyr		GGC Gly	570 GGA G1Y	9	CTC		GCC Ala	FIGURE
	TCC		ACG Thr	470	CGC Arg	520	GCC	GTT Val		AAT Asn		TAT Tyr	
	GCT	420	GCG Ala	47	CTC		CTC	CTA		CAG	* 099	CCA	
370	GAC	7'	AAC Asn		TGC		GAT Asp	50 GTG Val	610	GTT Val	v	ATT Ile	
	TTC Phe		TTC Phe		GAC GAT ASP ASP	510	GCC	560 GGA G1 Gly Va		$_{\rm GGG}$		TTC	
	CGC Arg	410	GGC Gly	460		۵,	GAC	GCC Ala		GAC	650	TTT Phe	
360	GAC	4	CGT	1	CTC		GAA Glu	AGG Arg	*	TCT	9	CCG	
. •	ATC Ile		ATC Ile		CGG Arg	200	CTC	550 GAG Glu	•	TTC Phe		TCC	
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	ACT		ATC	GCG Ala		TCT Ser	930	GAC Asp		GAG Glu		TAT Tyr
	TCA		CAT His	GCT Ala	880	TTA	ഗ	AAG Lys		ATG Met		GAA Glu
	ATT Ile	780	AAT Asn	830 GAG G(Glu A.		GCT Ala		GAT Asp	•	TTG GTT . Leu Val	1020	GCA
730	TCG	-	GCC	ACT		AGG Arg	920	TGG Trp	970		Ä	ATT Ile
	TAT Tyr		GCC Ala	GGA G1y	870	TGC	9	CCG		GTA Val		ATT Ile
	AAC Asn	770	GCT Ala	820 GGA G1y	w	GCC		AGG Arg		GGA Gly	10	CCG
720	CCA	7.2	TAT Tyr	GCT		GTT Val		TCA	960	GCT Ala	1010	GCG Ala
	GGC Gly		$ ext{TTT}$	ATT Ile	860	TTC	910	GCC Ala		$_{\rm GGG}$		GGA Gly
	ATG		TGC	810 CTG ATG	æ	GGA Gly		ACT		GAA Glu		CGG
0	CTG	760	TAC	CTG Leu		GGA Gly		GAT CCT CAG ASP Pro Gln	950	GGT Gly	1000	AAA Lys
710	GGT		AAC Asn	GAC Asp		TTA	006	CCT	9	ATG		GCA ATG Ala Met
	TTG		TCC	800 GAG GCT Glu Ala	850	GGT Gly	٥,			GTG Val		GCA Ala
	GAT	750	ACT	80 GAG Glu		ATT Ile		gat Asp		TTT Phe	066	CAT His
700	ATC Ile		GCT Ala	GGT Gly		CCA	068	AAT Asn	940	GGC Gly		GAG Glu
	GCC		$_{ m Cys}$	cga Arg	840	* ATT Ile	æ	AGG		GAT		TTG Leu
	CTT Leu	740	GCA Ala	790 CGC Arg	w	GTC Val		CAA		CGT Arg	980	AGC

FIGURE 6 3/5

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AGG		GAT Asp	1170	ACT		GTT Val		ATC Ile	ATT Ile		AAT
OCCA Pro	1120	GAA Glu	7	GCG Ala		AAA Lys		ATG Met	.0 ACC Thr	1360	TTT Phe
1070 GAT CCA ASP Pro	П	CIC		CAT		AAG Lys	1260	TCA	1310 GCA ACC Ala Thr	-	CAA TTT Gln Phe
ACT		AGT	0.0	GCT Ala	1210	ATT AAG	12	AAG Lys	ATC		AAT Asn
ATG Met	1110	AGC Ser	1160	ATA AAT GCT Ile Asn Ala	П	GCC Ala		ACT Thr	GCC Ala	1350	CTT CAT CCC AGC ATT Leu His Pro Ser Ile
1060 TAT CAT 1 Tyr His 1	11	GAG Glu		ATA Ile		GAG ATA AAT Glu Ile Asn	0.9	GCA	1300 CTT GAA Leu Glu	13	AGC
TAT TYF		ATT Ile		AAT TAC A	1200	ATA Ile	1250	AAT Asn			CCC
3CT Ala	0.0	TGC	1150	AAT Asn	17	GAG Glu		ATC Ile	GGT Gly	0 1	CAT His
E .O.	1100	TCG		GTC Val		GCC Ala		AAA Lys	1290 TCA GGA GGT Ser Gly Gly	1340	
1050 TGT GAS Cys Asi		TCC		GAG Glu	0	GAT CTT Asp Leu	1240	ATC Ile	12 TCA Ser		TGG
AAC Asn		GTC Val	1140	CCT GAA Pro Glu	1190	GAT Asp	-	GAA Glu	GCA Ala		GGC Gly
1040 GCA GTC Ala Val	1090	GGT Gly	11			666 G1y		AAG Lys	1280 CTT GGA Leu Gly	1330	ACC
1040 GCA G1 Ala Ve	П	CTT		TCA		GCT Ala	1230	ACC	1280 CTT G Leu G		ACC Thr
GGT Gly		GGG G1y	0 20	GGG GTC Gly Val	1180	CTT	12	AAC Asn	TGT Cys		ATA Ile
GGA G1y	1080	GAT Asp	1130		-	ACT Thr		AAG Lys	CAC His	1320	GGA Gly
1030 TTG Leu	10	GCT		GCC		TCT Ser	1220	TTC	1270 GGA Gly	E) El	AAG Lys

FIGURE 6

1410	CAG CAA Gln Gln		. GGG CAC	1510	ACTTGGTTCA	1570	TAAATGCCTT	1630	AGCCATTTAG	1690	СТСТGАТТТА	1750	GTTATTTAAG		CT
1400	AAC AAA AAG Asn Lys Lys	1450	GGA TTT GGA Gly Phe Gly	1500	TGA ATTCT	1560	AGCAATTTT	1620	GAATAGGTCG GTCCTTTGAT AGTTCCTCGA AGCCATTTAG	1680	TAAATCTAGT	1740	TGTTGTCAAT GTTATTTAAG	1800	GCTCTAGAGG ATCCAGCTTA
1390	ACT GTT GCC Thr Val Ala	1440	AAT TCT TTT Asn Ser Phe	1490	TTC AAG CCA Phe Lys Pro	1550	CAACTTGCAG	1610	GTCCTTTGAT	1670	ATTCCCATTT	1730	GTCATGTTTG	1790	GCTCTAGAGG
	TTC AAC Phe Asn	1430	ATC TCG Ile Ser	1480	TCA GCT Ser Ala	1540	GATAGGGCTT	1600	GAATAGGTCG	1660	ATCGAAGATG	1720	AAGATTTTGT	1780	AAGGGCGCCC
1380	TCG GTG GAC		ASD VAL ALA	1470	GTG GCA TTC Val Ala Phe	1530	AAATGCACAC CAGTTGCTGA GATAGGGCTT CAACTTGCAG AGCAATTTTT TAAATGCCTT	1590	CGTAATACCG	1650	TACTGTAATA ATCGAAGATG	1710	TGTATTAGAA AGACCAATGA AAGATTTTGT GTCATGTTTG	1770	ATAAAGCAAA AAAAAAAAA AAGGGCGGCC
1370	CCC GAG CCA Pro Glu Pro	1420	CAT GAA GTG His Glu Val	1460 1	AAC TCG GTT Asn Ser Val	1520	AAATGCACAC	1580	GTCGGAAGAG	1640	GATGATGTTT	1700	TGTATTAGAA	1760	ATAAAGCAAA

FIGURE 6 5/5

Sequence Range: 1 to 2369

	ataaaagag	120	TACCATACC	180	rccttttct	230 TCT TCC Ser Ser>	280	ATG TCT Met Ser>	330	TCT CCT Ser Pro>		CCA CTA Pro Leu>
20	GTACGCCTGC AGGTACCGGT CCGGAATTCC CGGGTCGACC CACGCGTCCG CATAAAAGAG	110	CTTCGATTCA TTACCATACC	170	CCCAAAGGGT ATCCTTTTCT	GCC GCC Ala Ala	270	GCC GCC TGC Ala Ala Ala Cys 1	320	TCC ATC TCC Ser Ser 11e Ser 3	370	CAA TGC GCC Gln Cys Ala
40	CGGGTCGACC	100	AGAGAGAGG ATCCATCGAA TGCGGCCACC CTCCTTTCAT	160	GGTCTTTCAT	CCTCCA AT	260	TGG CTC CTT Trp Leu Leu	310	CTT CCG CCT Leu Pro Pro	360	ATT CTC TCC Ile Leu Ser
30	CCGGAATTCC	06	TGCGGCCACC	150	GCCTTTTCCG	210 CAGTCAGTTC	26	TGT ACG Cys Thr		GAC CCT ASP Pro	350	cGC CGG Arg Arg
20	AGGTACCGGT	80	ATCCATCGAA	140	TCCATTTTCC	200 CTCAAAGGGT	250	TCC CCT CTC Ser Pro Leu	300	CAC CCC TCC His Pro Ser	340	CTC TCC CGC Leu Ser Arg
10	GTACGCCTGC	70	AGAGAGAGGG	130	ATTCCGCTGA	190 ATCCTATCTT	240	CTG CTC GCT Leu Leu Ala	290	ACC TCC TTC Thr Ser Phe	8	CGC CGA CGC Arg Arg Arg

FIGURE 7

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420	ACC CTC GTC Thr Leu Val>	470 TAT ACA TCC Tyr Thr Ser>	520	AGG CAC CGG Arg His Arg>	570	GTG GCT CTG Val Ala Leu>	0	ATC AAA CAG Ile Lys Gln>	099	CTA GGC CAT Leu Gly His>	710 AGT GGC ATA Ser Gly Ile>
	CAT	TAC		CGC	260	GCC	610	AGT		CCT	ACG Thr
	TTC	50 GAC ASP	510	ACC Thr	Ŋ	ATG Met		CCA Pro		ACT	700 GAT GGA ASP Gly
410	AGT Ser	460 CAT GA His As		ACC		GCA Ala		AAG Lys	650	GTG Val	70 GAT ASP
4	TCC	TGC Cys		CGC	20	GAG Glu	600	AAG Lys	•	GTG Val	CTT Leu
	GGA	CCC	200	ATT Ile	5	AGG		AAG Lys		GGT Gly	CTG
0	CGC	450 GAG Glu	u	CCC		TCC		ACA Thr	640	ATG Met	690 AAT Asn
400	CTC	TTC		AGA Arg		CCT	290	ACC	ý	GGA Gly	AAT Asn
	GCC	TGC	490	TCC	540	TCC		GTT Val		ACT	TAC
	TCC Ser	440 GCC Ala	4	GGA Gly		GCT		GAA Glu		GTG Val	680 GTT TTC Val Phe
390	TCC	CTC Leu		TTC		CGA Arg	580	cag Gln	630	GTT Val	
	GCT	TAC		TTG	530	AAT Asn	28	GAA Glu		GTA Val	gat Asp
	TCT	10 TCT Ser	480	TCC Ser	U ,	CTC		CCT		CGA Arg	670 GAC CCT ASP Pro
380	CCT	430 ACC TO Thr Se		GCA		AGG Arg		CAA Gln	620	CGG	67 GAC ASP

FIGURE 7

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GCT Ala>	810	CTC Leu>		AAG Lys>		CTA Leu>	ATG Met>	00	AAG Lys>	.050	GCT Ala>
		AAG Lys		GGC Gly		GAG Glu	₫ 5-	100	AAG Lys		TCA
		CCG Pro	20	GCT Ala	006	AAA Lys	SGT Gly		TAT Tyr		GGA Gly
ACG	300		ά						TCA	040	ATG Met
CCT	~	GTG Val		CTG		GTG Val	10 GCA Ala	990		1(AAT ATG Asn Met
TTT Phe		TGG		ATG Met	390		9. TCA Ser		agg Arg		ACA Thr
CAA Gln	90	GGT Gly	840		~	GAA Glu	GGC Gly		CTA	0 8	GCT ACC Ala Thr
GCT	7.5	GAT Asp		CTA		ACC Thr	ATT Ile	086	GCC Ala	103	GCT Ala
		ACA Thr		ATG Met	30	ATC Ile	930 CTC Leu	01	GAA Glu		TTC
GAT Asp			330	TTC Phe	88	GGA Gly	GTT Val		ATT Ile		CCT
TTT Phe	780	TTC Phe	w	AAG Lys		GGT Gly	GGA Gly	0,	GCC Ala	.020	GTA Val
ACC		TCT		GAC		GAT Asp	J20 TGC Cys	97	GAT Asp	-	TGT Cys
GAG Glu		AAG Lys	0.	ATG Met	870	ACA Thr	aaa Lys		AAT Asn		TTT Phe
ATA Ile	071	ATC Ile	8	AGG Arg		TTA	AGA Arg			110	CCC
		GAG Glu					LO AAA Lys	960		10	AAT Asn
AGC		GGA Gly		TCT	860	AAA Lys	91 GAT ASP		AAG Lys		ATG Met
	GAG ATA GAG ACC TTT GAT TGT GCT CAA TTT CCT ACG AGA ATT Glu ile Glu Thr Phe Asp Cys Ala Gln Phe Pro Thr Arg Ile	GAG ATA GAG ACC TTT GAT TGT GCT CAA TTT CCT ACG AGA ATT Glu Ile Glu Thr Phe Asp Cys Ala Gln Phe Pro Thr Arg Ile 770 780 800	Gad ATA GAG ACC TTT GAT TGT GCT CAA TTT CCT ACG AGA ATT Glu Ile Glu Thr Phe Asp Cys Ala Gln Phe Pro Thr Arg Ile 770	Gad ATA GAG ACC TTT GAT TGT GCT CAA TTT CCT ACG AGA ATT Glu Ile Glu Thr Phe Asp Cys Ala Gln Phe Pro Thr Arg Ile 770 GAG ATC AAG TCT TTC ACA GAT GGT TGG GTG GCC CCG AAG GIU Ile Lys Ser Phe Ser Thr Asp Gly Trp Val Ala Pro Lys Agg S20 820 820 840 850	Gag ATA GAG ACC TTT GAT TGT GCT CAA TTT CCT ACG AGA ATT Glu Ile Glu Thr Phe Asp Cys Ala Gln Phe Pro Thr Arg Ile Glu Ile Lys Ser Phe Ser Thr Asp Gly Trp Val Ala Pro Lys Aga AGG AGG AGG AGG AGG ATC ACG ACG ACG GLU Ile Lys Ser Phe Ser Thr Asp Gly Trp Val Ala Pro Lys Aga AGG AGG AGG AGG AGG AGG AGG AGG ATG ACG AGG AGG AGG AGG AGG AGG AGG AGG AG	C GAG ATA GAG ACC TTT GAT TGT GCT CAA TTT CCT ACG AGA ATT TG Glu Ile Glu Thr Phe Asp Cys Ala Gln Phe Pro Thr Arg Ile Ile Glu Thr Phe Asp Cys Ala Gln Phe Pro Thr Arg Ile Ile Glu Thr TrC TrC Aca GAT GGT TrG Gr Gr Gr Gr Acg AAG Y TrC Aca GAT GGT TrG Gr G	C GAG ATA GAG ACC TTT GAT TGT GCT CAA TTT CCT ACG AGA ATT CGL GLU Thr Phe ASP Cys Ala Gln Phe Pro Thr Arg Ile Su TTT CT TTC TCC ACA GAT GTT TTP CTT ASP GLY TTP Val ALA SP GLY TTP ASP GLY TTP Thr ASP GLY TTP CTC ACA GAT GTT TTP Thr ASP GLY TTP Val ALA SP GLY TTP Thr ASP GLY THR	C GAG ATA GAC TGT GCT CAA TTT CCT AGG AGA ATA AGG AGG	C GAG ATA GAG ACC TTT GAT TGT GCT CAA TTT CCT ACG AGA ATT ACA GAL GAL GAL GAL GAL GAL GAL GAL GAL GA	C GAG ATA GAG ACC TTT GAT TGT GCT CAA TTT CCT ACG AGA ATT TC GLU IIe Glu Thr Phe Asp Cys Ala Gln Phe Pro Thr Arg IIe Glu IIe Glu Thr Phe Asp Cys Ala Gln Phe Pro Thr Arg IIe Glu IIe Lys Ser Tr TC ACC ACA GAT GGT Tr	C GAG ATA GAG ACC TTT GAT TGT GCT CAA TTT CCT ACG AGA ATTT CGT TTT TT

FIGURE 7

060 1080 1090 1090 **	A ATG GAC TTG GGA TGG ATG GGG CCC AAC TAC TCG ATA TCT A Met Asp Leu Gly Trp Met Gly Pro Asn Tyr Ser Ile Ser>	1110 1120 1130 1140.	IT GCA ACG AGT AAC TTT TGT ATA ATG AAT GCT GCG AAC CAT IS Ala Thr Ser Asn Phe Cys Ile Met Asn Ala Asn His>	1160 1170 1180 1190 3A GGC GAA GCA GAT GTG ATG CTT TGC GGG GGC TCA GAT GCG cg Gly Glu Ala Asp Val Met Leu Cys Gly Gly Ser Asp Ala>	1210 1220 1230 1240	IA CCT ATT GGT ATG GGA GGT TTT GTT GCA TGC CGA GCT TTG le Pro Ile Gly Met Gly Gly Phe Val Ala Cys Arg Ala Leu>	0 1260 1270 1280 1290	* GA AAT TCC GAC CCT ACT AAA GCT TCA AGA CCA TGG GAC AGT rg Asn Ser Asp Pro Thr Lys Ala Ser Arg Pro Trp Asp Ser>	1300 1310 1320 1330	AT GGA TTT GTT ATG GGG GAA GGA GCT GGA GTG CTA CTA CTA sp Gly Phe Val Met Gly Glu Gly Ala Gly Val Leu Leu>	1350 1360 1370 1380	TG GAG CAT GCA AAG AAA AGA GGT GCG ACT ATT TAC GCA GAA eu Glu His Ala Lys Lys Arg Gly Ala Thr Ile Tyr Ala Glu>	FIGURE 7
1060	ATG CTT GCA ATG GA Met Leu Ala Met As	1100 1110	r GCT TGT GCA r Ala Cys Ala	1150 ATA ATC AGA GGC GA Ile Ile Arg Gly Gl	1200		1250		1300		1340 1350		

SUBSTITUTE SHEET (RULE 26)

0.	GCT Ala>	.530	GCC Ala>		CAC His>		ATG Met>	GTA Val>	50	GAA Glu>	
148	TTG	***	CAT His		ATC Ile		TCA	570 TCA Ser	17:	TTG	
	GCT		GCC Ala	0	CTT Leu	,620 *	AAA Lys	16 GTT Val		AAT Asn	
	AAG Lys	20	AAT Asn	157	GCT	-	ACC Thr	GCA Ala		ATT Ile	
470	GAG Glu	15	ATA Ile		CAA Gln		TCA	50 GAA Glu	1710	AAT Asn	
П	ATA Ile		TAC Tyr		TAC	010	AAT Asn	166 GTG Val	,,	CCG	
	TGC	0	AAT Asn	.560	GAG Glu	16		GGT Gly		CAT His	
09	CTC	151	GTA Val	П	AAA Lys		AAA Lys	GGT Gly	002	ATC Ile	
14	ATT Ile		GAC		ATC Ile	0.0	TTA Leu	1650 GCC Ala	H	TGG Trp	
	GTG Val		GAA Glu	150	GAT Asp	160	GAG Glu	GCA		666 61y	
0	GGA Gly	500	AGG Arg	7	GGA Gly		AGA Arg	GGA Gly	06	ACT Thr	
145	GCT Ala	П	TCT		GCT Ala		AAC Asn	540 CTC Leu	169	AGG Arg	
	GGA Gly		src /al	0,1	CCG	1590		16 CTT Leu		ATA Ile	
	GAT Asp	06	GGA Gly	154	ACT Thr			CAC His		GCA Ala	
440	CCT	14	TCA		TCC			30 GGT Gly	1680	cAG Gln	
Н	CAC His		CAG Gln		ACA	1580	TGT Cys	162 ATT Ile	1 -1	GTT Val	
	1440 1450 1460 1470 1480	.440 * CCT GAT GGA GTG ATT CTC TGC ATA GAG AAG GCT TTG Pro Asp Gly Ala Gly Val Ile Leu Cys Ile Glu Lys Ala Leu	440 440 CCT GAT GGA GTG ATT CTC TGC ATA GAG AAG GCT TTG Pro Asp Gly Ala Gly Val Ile Leu Cys Ile Glu Lys Ala Leu 1490 1500 1500 1510 1610	440 1450 1460 1470 148 CCT GAT GCT GGA GTG ATT CTC TGC ATA GAG AAG GCT TTG Pro Asp GIY Val I16 Leu CYS I16 GIU LYS Ala Leu TCA GGA GTC TCT GGA GAA GAA GAA GCA TA TCA GGA GTC TCT AGG GAA GAA AAT TAC ATA AAT AAT	1450	440 1450 1460 1470 1480 1880 1480 1880	1440	1440	1440	1440	1440

FIGURE 7 5/7

FIGURE 7 6/7

1770	AAG Lys Lys>		rrr GGr Phe Gly>	1870	GTTTCCGTGT	1930	GTTGGTAGCT	1990	GAACCATGAC	2050	GTAGAGCAAT	2110	GTTGTACTTT	2170	CACGTAGTAA
1760	GTG GGT CCT Val Gly Pro	1810	TCA TTT GGG Ser Phe Gly	1860	ATC TAG GAC Ile ***>	1920		1980	GCTTTAGTCG GAACCATGAC	2040	AGAATTGTTG	2100	TITITCTCTG AAATCTCCCT CCTTGCAATA	2160	TTAACTCGGG
1750	AAA TTG CTC CLys Leu Leu	1800	TTG TCT AAT CLeu Ser Asn	1850	GCC CCT TAC Ala Pro Tyr	1910	AGTTTTGAGG	1970	TGTGTCCGGA	2030	ACTCCTTGCT	2090	AAATCTCCCT	2150	AACAAAGCTG
	GAT ACA Asp Thr	1790	GTC GGT Val Gly	1840	CTC TTC Leu Phe	1900	rcaaagctga 7	1960	CCATGAGTTT TGTGTCCGGA	2020	CACTTGATAT	2080	TTTTTCTCTG	2140	TGAAGAAGAG
1740	GGC		AAC GTT AAG Asn Val Lys	1830	AAC TCG TCC ATA Asn Ser Ser Ile	1890	GTGGAATTCT ACTCAACATA TCAAAGCTGA AGTTTTGAGG ACTCCAGCAT	1950	CCTTACGTCT CTAGACATGC	2010	GGATTGAGTA CTCATGGCGA CACTTGATAT	2070	TCATATTTT	2130	CGAGCTTTTC ATCGAGTCAG TGAAGAAGAG AACAAAGCTG TTAACTCGGG CACGTAGTAA
1730	AAC CCA GAT GAA Asn Pro Asp Glu	1780	GAG AGA CTG Glu Arg Leu		G CAC	1880	GTGGAATTCT	1940	CCTTACGTCT	2000	GGATTGAGTA	2060	ATTCATTATC	2120	CGAGCTTTTC

SUBSTITUTE SHEET (RULE 26)

2230	CCATTIGCCC TITGITITGC TCTCTATITC ATCACCGITT TGTGGTTTTA AAATTIGTAA	2290	AACTAGAAGA CTGGTTTAGA TTGGTTTGTT TTCTCATTGA TAATTGGGGR ATGTATGTTT	2350	TGGAAATAAA AAAAAAAA AAAAAAAA AAAAAAAA AAAAAA	
2220	TGTGGTTTTA	2280	TAATTGGGGR	2340	AAAAAAAA	
2210	ATCACCGTTT	2270	TTCTCATTGA	2330	AAAAAAAAA	
2200	TCTCTATTTC	2260	TTGGTTTGTT	2320	AAAAAAAAA	
2190	TTTGTTTTGC	2250	CTGGTTTAGA	2310	AAAAAAAAA	CTCTAGAGG
2180	CCATTTGCCC	2240	AACTAGAAGA	2300	TGGAAATAAA	2360 AGGCGGCCG CTCTAGAGG

FIGURE 7

1 to 2374
CGGAATTCCC GGGTCGACCC ACGCGTCCGC GACGCCAACC CACACAAAC
80
TTCCTCAGCT TCTCTTCA AGACGGACGC CATTGGCÂGC AGACAGACAG ACAGACAGAC
140
CCATAAAAGA GAGAGAGAG GATCCATCGA ATGCGGCCAC CCTCCTTTCA TCTTCGATTC
200
ATTACCATAC CATTCCGCTG ATCCATTTTC CGCCTTTTCC GGGTCTTTCA TCCCAAAGGG
260
TATCCTTTTC TATCCTATCT TCTCAAAGGG TCAGTCAGTT CCCTCCAATG CCTGCCGCCT
320
CINCCIGCT CGCINCCCCN CICIGIACGI GGCICCINGC CGCCIGCAIG ICTACCICCI
380
TCCACCCCTC CGACCCTCTT CCGCCTTCCA TCTCCTCTCC
440
GCCGGATTCT CTCCCAATGC GCCCCACTAC CTTCTGCTTC CTCCGCCCTC CGCGGATCCA

FIGURE 8 1/5

540 *	GACTACTATA	* 009	CGGAGGCTCA	*	TGCAACCTGA ACAGGAAGTT	720	TIGIGACIGG AAIGGGIGIG	780	TTTCTACAAT AATCTGCTTG ATGGAACGAG	840	CCTACGAGAA TTGCTGGAGA	006	GGATGGACAA	* 096	GAATCACCGA	1020	AGATGTGATG AAAGAGCTAG ATAAAAGAAA ATGCGGAGTT CTCATTGGCT CAGCAATGGG
530	CCTGCTTCGA GCCCTGCCAT GACTACTATA	590	CCGCAGGCAC	650	TGCAACCTGA	710	TTGTGACTGG	770	AATCTGCTTG	830		890	CTCTCTAAGA	950	ACAGATGGTG	1010	CTCATTGGCT
520	CCTGCTTCGA	580	TTCGCACCAC	640	весетвесте	700	CGGCGAGTAG	760		820	TGCTCAATTT	880	GGCCCCGAAG	940	GAAAGCATTA	1000	ATGCGGAGTT
510	TCTTACCTCG	570	TCCAGACCCA	630	ATCGAGCTTC CCCTTCCAGG GGAGGCAATG	069	ACCACAAAGA AGAAGCCAAG TATCAAACAG CGGCGAGTAG	750	TAGGCCATGA ACCTGATGTT	810	GAGATAGAGA CCTTTGATTG	870	ATGGTTGGGT	930	GTTCATGCTA TACATGCTGA CTGCTGGCAA GAAAGCATTA	066	ATAAAAGAAA
200	CCTCGTCACC	260	CTTGTTCGGA TCCAGACCCA	. 620	CCCTTCCAGG	089	AGAAGCCAAG	740	TAGGCCATGA	800		860	GATCAAGTCT TTCTCCACAG	920	TACATGCTGA	980	AAAGAGCTAG
490	GTTTCCATAC	550	CATCCGCATC	610	ATCGAGCTTC	670	ACCACAAAGA	730	GTGACTCCTC	790	TGGCATAAGC	850	GATCAAGTCT	910	GTTCATGCTA	970	AGATGTGATG

FIGURE 8

TGCGACTATT TACGCAGAAT TTCTAGGTGG GAGTTTCACT TGCGATGCCT ACCACATGAC	TGCGATGCCT	GAGTTTCACT	TTCTAGGTGG	TACGCAGAAT	TGCGACTATT
1500	1490	1480	1470	1460	1450
TATGGGGGAA GGAGCTGGAG TGCTACTACT AGAGGAGTTG GAGCATGCAA AGAAAAGAGG	GAGCATGCAA	AGAGGAGTTG	TGCTACTACT	GGAGCTGGAG	TATGGGGGAA
1440	1430	1420	1410	1400	1390
GAGAAATTCC GACCCTACTA AAGCTTCAAG ACCATGGGAC AGTAATCGTG ATGGATTTGT	AGTAATCGTG	ACCATGGGAC	AAGCTTCAAG	GACCCTACTA	GAGAAATTCC
1380	1370	1360	1350	1340	1330
CTTTGTCCCA	GCATGCCGAG	AGGTTTTGTT	TTGGTATGGG	AGATGCGGTA ATCATACCTA TTGGTATGGG AGGTTTTGTT GCATGCCGAG	AGATGCGGTA
1320	1310	1300	1290	1280	1270
GCGGGGGCTC	GTGATGCTTT	CGAAGCAGAT	TAATCAGAGG	AATGAATGCT GCGAACCATA TAATCAGAGG CGAAGCAGAT GTGATGCTTT	AATGAATGCT
1260	1250	1240	1230	1220	1210
ACTTTTGTAT	GCAACGAGTA	TACTGCTTGT	ACTCGATATC	SGGATGGATG GGGCCCAACT ACTCGATATC TACTGCTTGT GCAACGAGTA ACTTTTGTAT	GGGATGGATG
1200	1190	1180	1170	1160	1150
CAATGGACTT	GCTATGCTTG	TATGGGATCA	CTACCACAAA	PCCCTTTTGT GTACCTTTCG CTACCACAAA TATGGGATCA GCTATGCTTG CAATGGACTT	rccttttgt
1140	1130	1120	1110	1100	1090
AGAAGATGAA	ATTTCATATA	AGCCCTAAGG	ATGCCATTGA	RGGAATGAAG GTATTCAATG ATGCCATTGA AGCCCTAAGG ATTTCATATA AGAAGATGAA	rggaatgaag
080T	1070	1060	1050	1040	1030

FIGURE 8

								•									
1560	TGGCTCAGTC	1620	CTCCGGCTGG	. 1680	AGTTAAAAGT	1740	TGGAAGCAGT	1800	TGGAAAACCC	1860	AGATGAAGGC GTGGATACAA AATTGCTCGT GGGTCCTAAG AAGGAGAGAC TGAACGTTAA	1920	TGGGCACAAC TCGTCCATAC TCTTCGCCCC	1980	TTACATCTAG GACGTTTCGT GTGTGGAATT CTACTCAACA TATCAAAGCT GAAGTTTTGA	2040	GGACTCCAGC ATGTTGGTAG CTCCTTACGT CTCTAGACAT GCCCATGAGT TTTGTGTCCG
1550	GAGAAGGCTT	1610	GCCACATCCA CTCCGGCTGG	1670	CAAAACAGAG AGTTAAAAGT	1730	GCCGGTGGTG	1790	GATCCATCCG AATATTAATT TGGAAAACCC	1850	AAGGAGAGAC	1910	TCGTCCATAC	1970	TATCAAAGCT	2030	GCCCATGAGT
1540	CTGGAGTGAT TCTCTGCATA GAGAAGGCTT TGGCTCAGTC	1600	AAATGCCCAT	1660	CTGTTTCGGC	1720	AAATCAATGA TTGGTCACCT TCTCGGAGCA GCCGGTGGTG TGGAAGCAGT	1780		1840	GGGTCCTAAG	1900		1960	CTACTCAACA	2020	CTCTAGACAT
1530	CTGGAGTGAT	1590		1650	CTCTTATCCA	1710	TTGGTCACCT	1770	CAGGCAATAA GGACTGGGTG	1830	AATTGCTCGT	1890	GGTCGGTTTG TCTAATTCAT TTGGGTTTGG	1950	GTGTGGAATT	2010	CTCCTTACGT
1520	CCTGATGGAG	1580	AGGGAAGACG	1640	GAGTACCAAG	1700	AAATCAATGA	1760	CAGGCAATAA	1820	GTGGATACAA	1880	TCTAATTCAT	1940	GACGTTTCGT	2000	ATGTTGGTAG
1510	CGAGCCTCAC	1570	AGGAGTCTCT AGGGAAGACG TAAATTACAT	1630	AGATATCAAA GAGTACCAAG CTCTTATCCA CTGTTTCGGC	1690	TAATTCAACC	1750	TTCAGTAGTT	1810	AGATGAAGGC	1870	GGTCGGTTTG	1930	TTACATCTAG	1990	GGACTCCAGC

FIGURE 8

2100	ATACTCCTTG	2160	TGAAATCTCC	2220	AGAACAAAGC	2280	TCATCACCGT	2340	TTTTCTCAAA	
2090	GAGCTTTAGT CGGAACCATG ACGGATTGAG TACTCATGGC GACACTTGAT ATACTCCTTG	2150	TTTTTTTCTC TGAAATCTCC	2210	CTCCTTGCAA TAGTTGTACT TTCGAGCTTT TCATCGAGTC AGTGAAGAAG AGAACAAAGC	2270	CCTTTGTTTT GCTCTCTATT TCATCACCGT	2330	TITGIGGITI IAAAAIITGI AAAACIAGAA GACIGGITTA GAITGGITIG	
2080	TACTCATGGC	2140	тстсататтт	2200	TCATCGAGTC	2260	CCTTTGTTTT	2320	GACTGGTTTA	ATCC
2070	ACGGATTGAG	2130	ATATTCATTA	2190	TTCGAGCTTT	2250	AACCATTTGC	2310	AAAACTAGAA	2370 GCTCTAGAGG
2060	CGGAACCATG	2120	CTAGAATTGT TGGTAGAGCA ATATTCATTA TCTCATATTT	2180	TAGTTGTACT	2240	TGTTAACTCG GGCACGTAGT AACCATTTGC	2300	TAAAATTTGT	2350 2350 2370 AAAAAAAAA AAGGGCGGCC GCTCTAGAGG ATCC
2050	GAGCTTTAGT	2110	CTAGAATTGT	2170	CTCCTTGCAA	2230	TGTTAACTCG	2290	TTTGTGGTTT	2350 AAAAAAAAA

FIGURE 8 5/5

Sequence Range: 1 to 1580

666 G1y>	100	TCG Ser>	150	GTG Val>		GAT Asp>		TCT Ser>	AAA Lys>	340	CGC Arg>
50 TCT Ser	7(CAT His		AGG Arg		GGT Gly		GGA Gly	290 GCT Ala	ň	ATC Ile
GCA Ala		CAG Gln		AAA Lys	190	TTG	240	ATT Ile	CTT		$\frac{\text{GGG}}{\text{Gly}}$
AAT Asn		ACT Thr	140	TCC	1.9	TCT Ser		TTA	GAT Asp		ACG Thr
40 GCG Ala	90	GCA Ala	П	GTC Val	٠	CAG Gln		AAA Lys	280 AAT GAT Asn Asp	330	CGA Arg
40 ATG GCG Met Ala		AGG Arg		TTT Phe		AGG Arg	230	TGC			GTC Val
GGG		AGA Arg	130	GAG Glu	180	GAC	.,	GGA Gly	TCA		ACT Thr
GCT	80	CTG	13	TCG		TCT		AGA Arg	GTC Val	320	TGG ATT Trp Ile
30 CGTT		GCC		TCC		GAT	220	AGT	270 CAA Gln		TGG Trp
\GT'T']		CCT		TCT	170	CAG Gln	7	GTG Val	CTT Leu		GAA Glu
20 GA G	70	GTT Val	120	GGA Gly	••	GTT Val		CTT	GCT Ala	310	GAT Asp
AAGAC		TCA		CGT Arg		GCC		AGG	260 CCA Pro	m	AAT Asn
10 20 30 CCTGAATCGG ATTCAAGAGA GAGTTTCGTT GCTGGG		TCT		TCT	160	AGT Ser	210	CCG	ATA Ile		ACC
10 366 7		GGT Gly	110	TCG	Ä	TGT Cys		TCG	GCT		GAC
BAATC	09	CTG Leu		TCA		TGC Cys		CGC	50 TCT Ser	300	GTC Val
CCTC		TTT Phe		ATT Ile		TTT Phe	200	TCT	250 GGT TC Gly Se		ATT Ile

FIGURE 9 1/5

										٨		٨	
390	TCA Ser>		GAT Asp>		GGC Gly>	TTG Leu>	280	GTC Val>	630	GTG Val>		GGA Gly>	
	GCA Ala		AAT Asn		TTC Phe	330 CCT Pro	28	TTA Leu		CTA Leu		CGG Arg	
	TTA	o.	GCA Ala	480	CTT Leu	5 AAT Asn		GGT Gly		ATT Ile	019	GAT Asp	
380	AAT Asn	430	gac Asp		GAC	AAG Lys		TTG	620	AAT	φ.	ACC Thr	
m	ACA Thr		GTA Val		GAG Glu	520 SC TGC AAA 1 Y CYS LYS	570	GTG Val	•	AAC Asn		TGG Trp	
	CTT Leu		CAG Gln	470	CCT	52 TGC Cys		TTT Phe		TTT Phe		GAC	
0	AGT	420	GCA Ala	4	점점	66C G1y		GGA Gly	610	GGT Gly	* 099	GTT Val	
370	GAT		ATG		TCT	CTT	260	AGT	9	GGG G1y		TAT	6
	AAA Lys		GAG Glu	460	ACT Thr	510 GCA Ala	5,	TGC		GGT Gly		CGG Arg	SURE 2/5
	$_{\rm GLY}^{\rm GGT}$	410	CTA	46	TGT	AAA Lys		GCA		AGA Arg	650	TCT	FIGURE 2/5
360	TCA	4	GCT		ATG Met	TCG	550	GCT	009	ATT		CTT	
	CTC		AAA Lys		TTG	500 ATA Ile	Ŋ	ACC Thr		CAC		TCT	
	GTT Val	400	AGG Arg	450	GTT Val	CAG Gln		ATT Ile		TGC Cys	640	GAT Asp	
350	AGG Arg	4	GCA Ala		ATG Met	CCT		GAC	590	GCT	9	GCT	
(*)	CGA Arg		GCA		GAT	490 T GCT Er Ala	540	TAC TYT		GCT		GGT Gly	
	AAC Asn		GAG Glu	440	GTG Val	49 AGT Ser		TCT		TCA		ATT Ile	

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	TCA Ser>	GAT Asp>	820	GTT Val>	870	AGG Arg>		CGC Arg>		AAG Lys>	GCA Ala>
	CAG Gln	770 AGC Ser	8	GAA Glu		CCA Pro		TTC		GGA G1y	cAG CAG Gln
720	GTG Val	CAT His	•	GAT Asp		CCA Pro	910	GTA Val	960	CTT	1010 CAT CAG
	GTG Val	TTG		GAA Glu	860	TTT Phe	91	GAG Glu		GC2 Ala	CTJ
	GTA Val	760 T GAT e Asp	810	AAA Lys		GAT Asp		AAA Lys		TCA	1000 TTG CTG
710	GCT	댸		ATC Ile		AGA Arg		GGT Gly	950	GAA Glu	1000 TTG C
•	GGA Gly	GCT Ala		GCA Ala	850	ATC Ile	900	AAC Asn	O1	ATC Ile	TGG Trp
	GCT	TTT Phe	800	GCT	ά	TCC		ATG Met		TCA	GAC Asp
700	GCT	750 CTC Leu	:	AAA Lys		GGG G1y		CAA Gln	940	CAG Gln	990 ATC Ile
7	GAT Asp	666 61y		CTA		AAT Asn	890	ATC Ile	76	CCT Pro	AAC Asn
	GGA Gly	GAT Asp	790	CAT His	840	CAT His	w	TGC		GTG Val	TCC
	TTT Phe	740 GAA Glu	7.9	AGG Arg		GGA		TCT		TCT	980 GGA Gly
069	CTC	GAG Glu		CAA Gln		CTG	880	TAC	930	CGC	, 9 AAT Asn
	ATT Ile	GCT		GGG Gly	830	GCC	88	TCA		TGC	CTT Leu
	TGT Cys	GAT ASP	780	GAT Asp	w	AAA Lys		TCT		GCT	o GGT Gly
680	ACA Thr	730 TGT G2 Cys As		GGA G1y		GAT Asp		CGT Arg	920	TTT Phe	970 GCC GGT Ala Gly

FIGURE 9 3/5

1060	CCT CAA Pro Gln>	1110	GCG GCA Ala Ala>		GTG AAG Val Lys>		ACA TGG Thr Trp>	1260	CACTGCAGCT	1320	AAGAAGTCAG	1380	TCGTTCCCCT
1050	CGT CTA GAG GTT Arg Leu Glu Val	1100	AAC ACT AGT Asn Thr Ser	1150	AGT GGA AAT Ser Gly Asn	1200	GCC GGA CTC Ala Gly Leu	1250	GGA TAA GACTGAA GCCGAGCCAG CACTGCAGCT	1310	CCANAAAAAG	1370	CTTCATCACA TTGCCCTTTT TCGTTCCCCT
	GTA GCA ACA CGT (Val Ala Thr Arg I	1090	AAT TAC GGG AAC Asn Tyr Gly Asn	1140	CTA GAC GAA GCT GTG AGG Leu Asp Glu Ala Val Arg	1190	rrr GGC Phe Gly	1240	TAA GACTGAA ***>	1300	GCTTCCATGA	1360	CTTCATCACA
1040	GCA Ala	**	TTG GCA Leu Ala	1130	GAC GAA G	1180	ACC GCA GGA Thr Ala Gly	1230	TGG Trp	1290	CGAAATTTT	1350	CGACACGAT
1030	* CAG AGG ATC ATT GAT Gln Arg Ile Ile Asp	1080	ATT ATC TCA AAC Ile Ile Ser Asn		CCC TTG GCA CTA Pro Leu Ala Leu	1170	GGT CAC GTG ATT GCA Gly His Val Ile Ala	1220	ATT ATC AGG Ile Ile Arg	1280	CCGATGITIC ACGAAAITIT GCTTCCAIGA	1340	TCTTTTATGG AGCAAGCAAC ACGACACGAT
1020	* AAT CAG AGG ASN Gln Arg	1070	GAA CGA ATT Glu Arg Ile	1120	TCC ATT CCC Ser Ile Pro	1160	CCG GGT CAC Pro Gly His	1210	GGT TCT GCT Gly Ser Ala		TCCTCTCAAA	1330	TCTTTTATGG

FIGURE 9

1430 1440 *	TITCCATTAG TITGATGATT TIGCTGACAA TACAATACCC ATAGTITCTT TIGTCCCCAA	1490 1500	TAAGTTATTT GTTTCTTGTT TAATTGTTCA GCTTTTACTT CATTTTGTCT CGGGACATTG	1550 1560	GAGATGACAG CATAAACATC ATGTTTATAT TTTGCTAAAA AAAAAAAAA AAAAAAAA	
	ATAG		CATT		AAAA	
3	TACAATACCC	1480	GCTTTTACTT	1540	TTTGCTAAAA	
1	TTGCTGACAA	1470	TAATTGTTCA	1530	ATGTTTATAT	
	TTTGATGATT	1460	GTTTCTTGTT	1520	CATAAACATC	1570 1580 AAAAAAAA AAAAAAAA
2	TTTCCATTAG	1450	TAAGTTATTT	1510	GAGATGACAG	1570 AAAAAAAAA

FIGURE 9 5/5

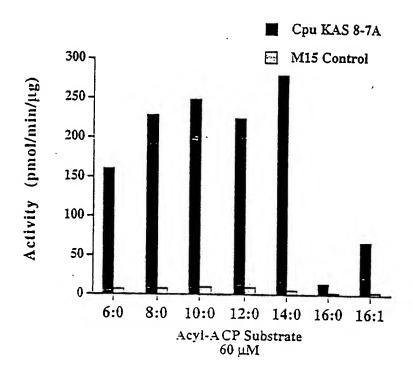


FIGURE 10

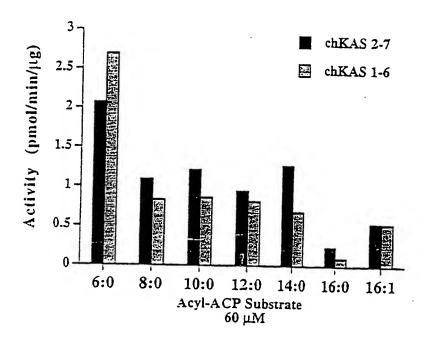


FIGURE 11

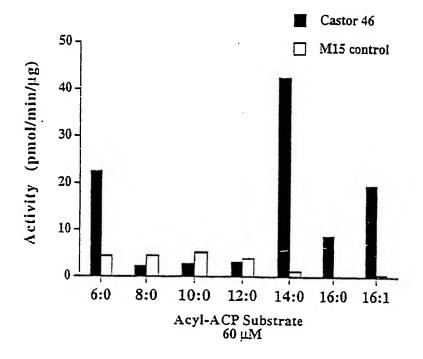
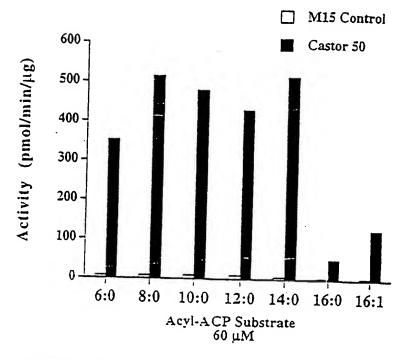


FIGURE 12



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FIGURE 13

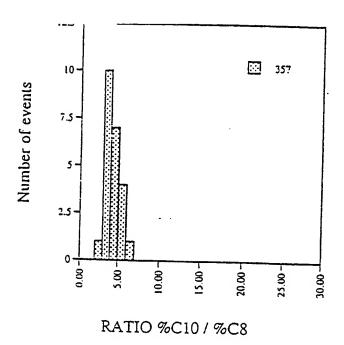


FIGURE 15

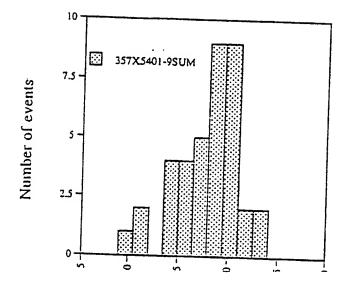
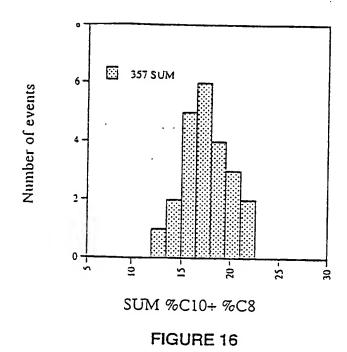


FIGURE 15 2/2



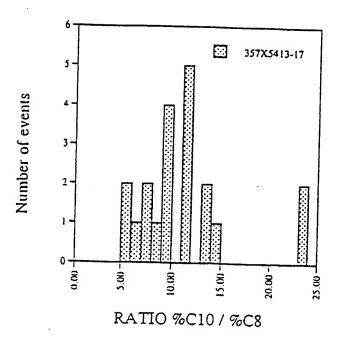


FIGURE 17 1/2

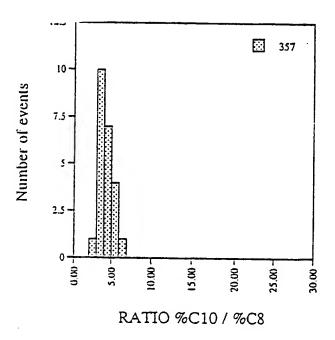


FIGURE 17

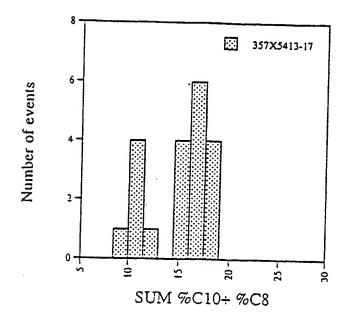
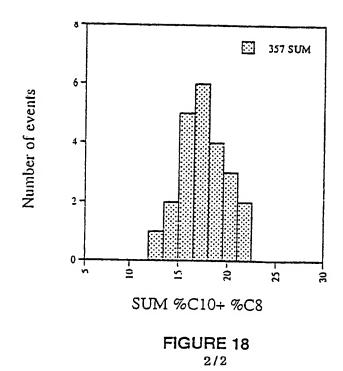
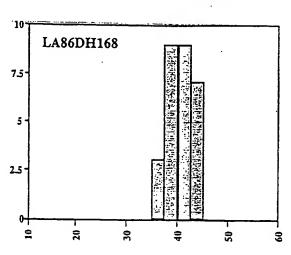


FIGURE 18 1/2







12:0 levels (w%)

FIGURE 19



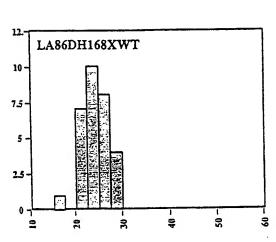
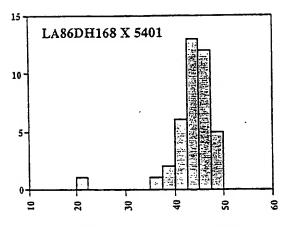


FIGURE 19
3/3
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12:0 levels (w%)

FIGURE 19 2/3

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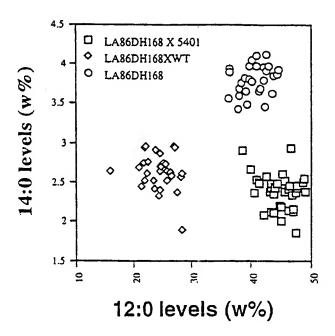
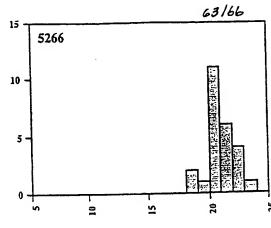


FIGURE 20

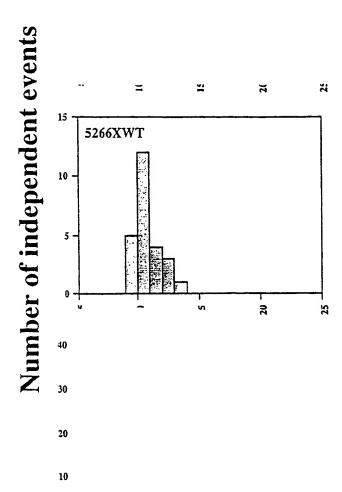




18:0 levels (w%)

FIGURE -21

1/3



18:0 levels (w%)

FIGURE 21. 2/3

Number of independent events

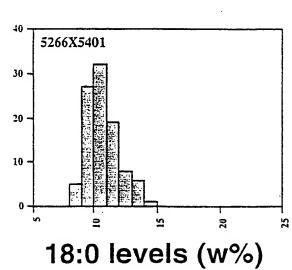


FIGURE 21 3/3

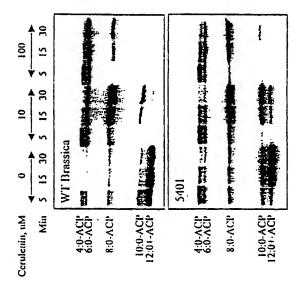


FIGURE 22

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